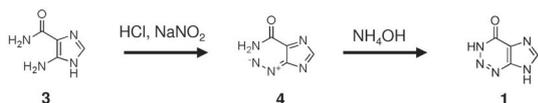


The Source of “Fairy Rings” :
2-Azahypoxanthine and its Metabolite Found in a
Novel Purine Metabolic Pathway in Plants

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URL	http://hdl.handle.net/10297/00026817

a) Chemical synthesis



b) Histidine metabolism
Purine metabolism

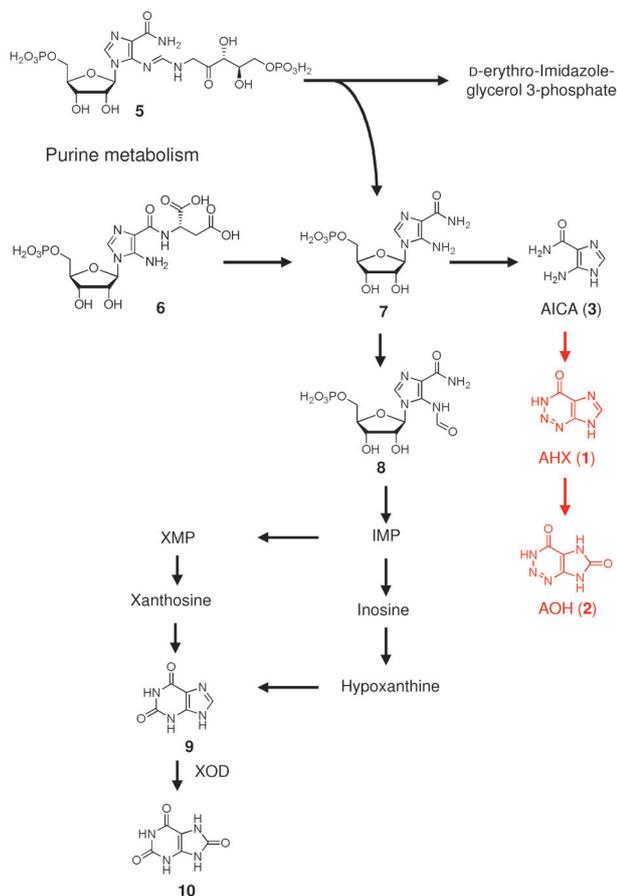
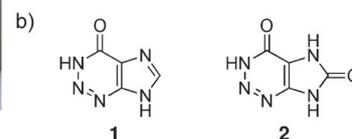
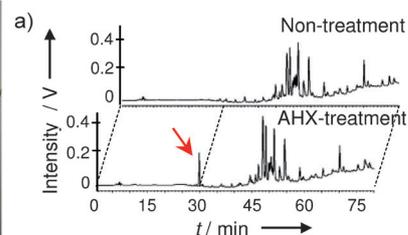


Figure 1. a) Synthetic route from AICA (3) to AHX (1). b) Purine metabolic pathway in animals, plants and microorganisms, including the novel route. The route in black in (b) was adapted from the KEGG (Kyoto Encyclopedia of Genes and Genomes). New metabolites and route are indicated in red. 1 = 2-azahypoxanthine, 2 = 2-aza-8-oxohypoxanthine, 3 = 5-aminoimidazole-4-carboxamide, 4 = 4-diazo-4H-imidazole-5-carboxamide, 5 = phosphoribulosylformimino-AICAR-phosphate, 6 = succino-AICAR, 7 = AICAR ribotide, 8 = 5-formyl AICAR, 9 = xanthine, 10 = uric acid, IMP = inosine monophosphate, XOD = xanthine oxidase, XMP = xanthine monophosphate.

tion of a novel compound, 2-aza-8-oxohypoxanthine (AOH; 2), as the metabolite, the structure of which was determined by X-ray crystal analysis (Figure 2b,c; see also Figure S1 in the Supporting Information). This conversion from 1 into 2 was also observed in *Arabidopsis*, tomato, and turfgrass (Figure S2), and reminded us of the reaction of xanthine oxidase (XOD) to produce 10 from 9 (Figure 1).

Therefore, 1 was treated with commercially available XOD, and 2 was quantitatively obtained (Figure S3). Compound 2 elongated the seedlings of bentgrass and rice in a fashion similar to 1 (Figures S4 and S5). In addition, 1 and 2



c)

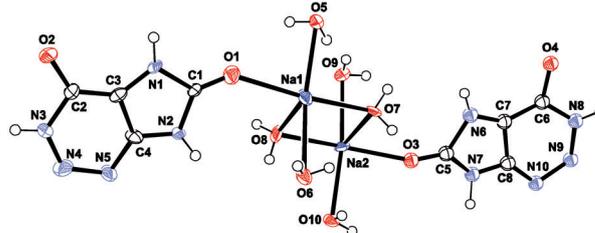
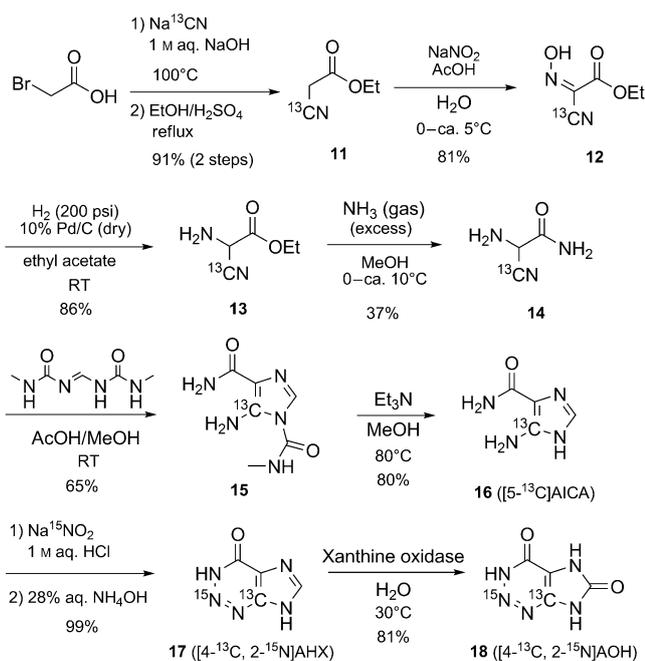


Figure 2. a) HPLC profile of the metabolite (2) of 1 in 1-treated rice shoots. Extracts of rice shoots were analyzed by reverse-phase HPLC using a Develosil C30-UG-5 column with a solvent gradient (2% MeOH in 0.05% trifluoroacetic acid (TFA) for 12 min, 2–100% MeOH in 0.05% TFA for 60 min, and 100% MeOH for 20 min) at a flow rate of 0.5 mL min⁻¹ at a wavelength of 254 nm. The arrow indicates the peak of 2 (29.6 min). b) Structures of AHX (1) and AOH (2). c) ORTEP drawing of 2 with ellipsoids set at 50% probability. Hydrogen atoms are shown as small spheres of arbitrary radii.

exhibited growth-yielding positive effects on rice. When rice was grown in soil supplemented with 1 or 2 (5 μM) in pot experiments, the seed yield per plant increased from 41.7 to 59.3 g (rate of increase, 42.2%) or 36.9 to 46.3 g (25.5%), respectively (Table S1).

To support the existence of 1 and 2 in rice, we quantified the endogenous 1 and 2 levels in rice cultivated under aseptic conditions by liquid chromatography/tandem mass spectrometry (LC-MS/MS) using [4-¹³C,2-¹⁵N]AHX (17) and [4-¹³C,2-¹⁵N]AOH (18) as internal standards (Scheme 1). We detected 1 (457 ± 75 ng kg⁻¹ F.W. in shoot and 273 ± 52 ng kg⁻¹ F.W. in root) and 2 (1289 ± 406 ng kg⁻¹ F.W. in shoot and not detected in root) (Figure S6). These results indicated that 1 and 2 are endogenously synthesized in both the shoot and root of rice. The amounts of endogenous 1 and 2 in rice were similar to those of the known plant hormones, strigolactones^[5] and brassinosteroids.^[6] Furthermore, 1 and/or 2 were detected in *Arabidopsis*, rice, bentgrass, zoysiagrass, tomato, potato, *Eucalyptus*, *Chlorella*, and *Parachlorella* (Figure 3; see also Table S2 and Figures S7 and S8).

To confirm the existence of our hypothetical biosynthetic route from 3 to 1 and 2 in plants (Figure 1), We performed a pulse chase labeling experiment using rice seedlings cultivated in liquid medium with [5-¹³C]AICA (16; 0.1 mM) and analyzed the incorporation rate of 16 into the seedlings, and the amount of [4-¹³C]AHX (19) and [4-¹³C]AOH (20) in



Scheme 1. Synthesis of isotope-labeled **1**, **2**, and **3**.

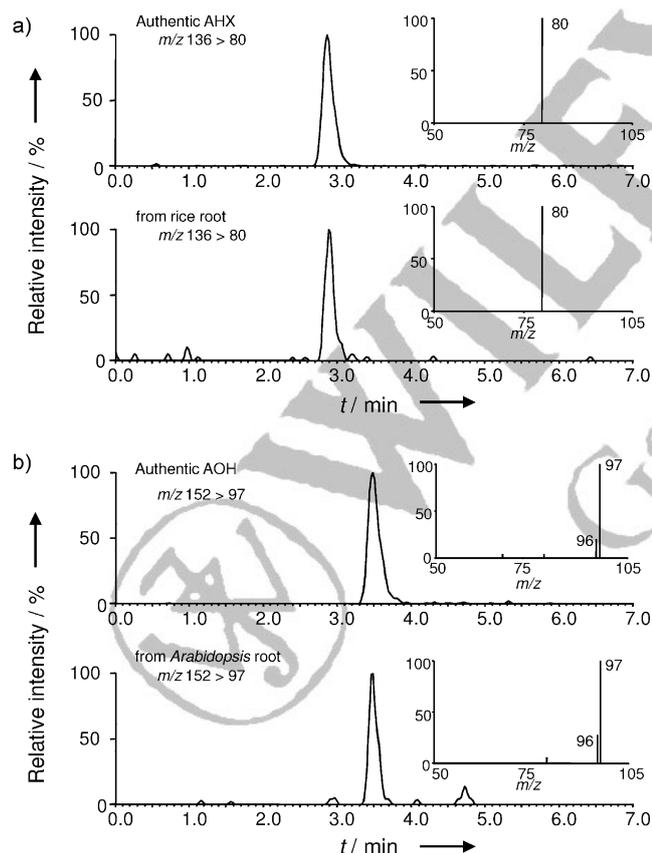


Figure 3. Identification of **1** and **2** in rice root (a) and *Arabidopsis* root (b) by negative mode of LC-MS/MS. a,b) LC-MS/MS chromatogram and MS/MS spectra of **1** and **2**, authentic sample (top), and root extracts (bottom). Characteristic transitions (precursor ion to daughter ion) for **1** and **2** were monitored.

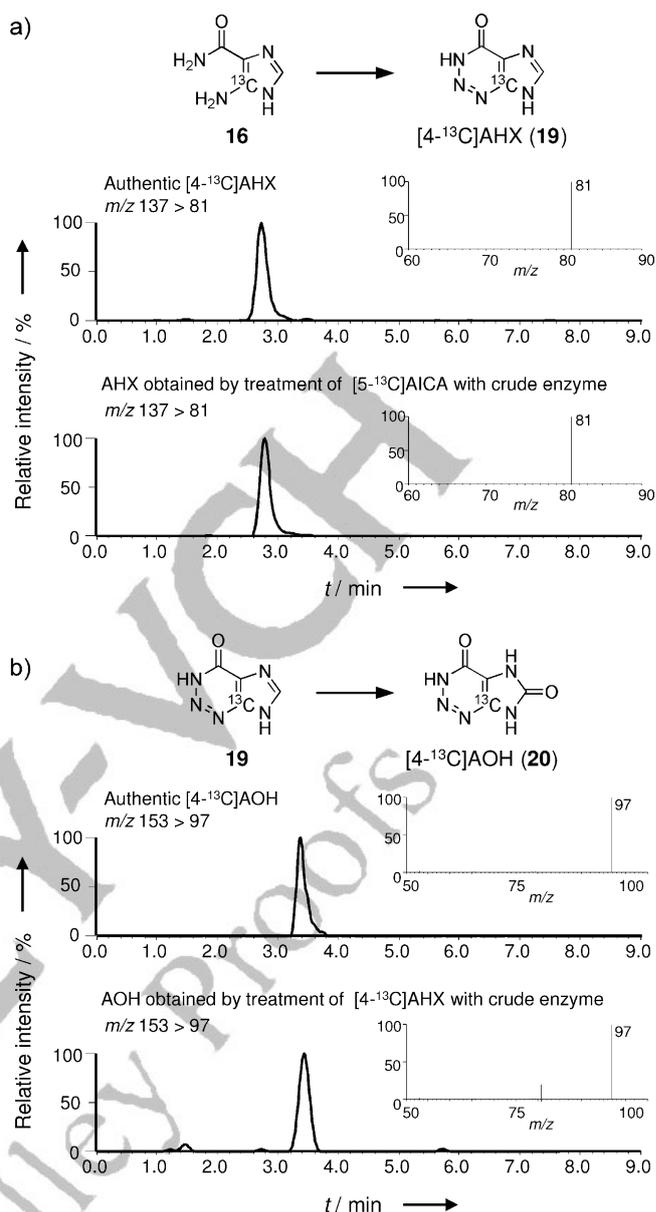


Figure 4. Detection of **19** (a) and **20** (b) in reaction mixtures treated with the crude rice enzymes. LC-MS/MS chromatogram and MS/MS spectra obtained for **19** and **20**, authentic sample (top), and root extracts (bottom). Characteristic transitions (precursor ion to daughter ion) for **19** and **20** were monitored.

the seedlings. A trace of **16** was detected with in the medium, and **20** was found in the seedlings. Compound **16** in the culture medium was incorporated at 99.98% into the rice seedlings, and this incorporated **16** was then converted into **20** through **19** (Figure S9).

Crude enzymes that catalyze the conversion from **16** into **19** and from **19** into **20** were extracted from rice and *Arabidopsis*, respectively, and fractionated by ammonium sulfate precipitation (Table S3). The supernatant obtained by ammonium sulfate precipitation of rice-root extracts, which showed the strongest activity of the reaction from **16** to produce **19**, was further separated by ultrafiltration, and the resulting fraction with a molecular weight of 10000–50000

exhibited enzymatic activity (Figure 4a). The fraction with molecular weight over 50000, which was obtained by ultra-filtration of the precipitates from a 30% saturation of ammonium sulfate showed enzyme activity for the conversion of **19** into **20** (Figure 4b).

All of these results lead to the conclusion that **1** and **2** are new metabolites in a novel purine metabolic pathway in plants, at least in rice and *Arabidopsis*. As mentioned above, all of the plants and algae analyzed contained **1** and/or **2**, thus indicating that the pathway is conserved in other organisms. In addition, the molecular mechanism of the growth-promoting activity of the new metabolite **2** was investigated using oligo DNA microarrays of rice (*Oryza sativa* L. cv. Nipponbare). The gene expression profile of rice treated with **2** was very similar to that of **1** (Figure S10). The most conspicuous results of the microarray analysis was significant induction of glutathione *S*-transferases (GST; Os09t0367700-01, Os03t0785900-01, Os10t0528300-01), an aquaporin, OsTIP2;1 (Os02t0658100-01), and Bowman-Birk type proteinase inhibitor (Os01t0127600-01) upon treatment with **1** or **2** (Figure S10). The up-regulation was also confirmed by reverse transcriptase-polymerase chain reaction (RT-PCR; Figure S10 and Table S4).^[3] These results suggest that the mysterious fairy that stimulates the plant growth in fairy rings might not only be **1**, but also **2** (Figure S4 and S5).

It will take some time before the new purine pathway and the mechanism of action of **1** and **2** are completely elucidated. However, from all the results it is clear that the chemical

“fairies” **1** and **2** are commonly biosynthesized by the novel purine metabolic pathway in plants.

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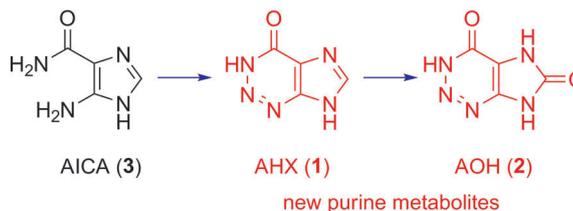
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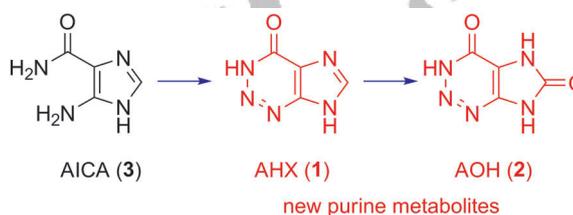
The fairy of the rings: The common metabolite, 2-aza-8-oxohypoxanthine (2), of the "fairy", 2-azahypoxanthine (1), was isolated from rice, thus supporting the existence of endogenous 1 and 2 in plants. Compound 1 is synthesized from 3 by treatment with NaNO₂ and then

NH₃; 1 can then be converted into 2 by xanthine oxidase. The new purine pathway to form the compounds occurs in rice and *Arabidopsis* through a route similar to the chemical synthesis. ■■ Revised text ok? ■■

Naturstoffe

J.-H. Choi, T. Ohnishi, Y. Yamakawa, S. Takeda, S. Sekiguchi, W. Maruyama, K. Yamashita, T. Suzuki, A. Morita, T. Ikka, R. Motohashi, Y. Kiriwa, H. Tobina, T. Asai, S. Tokuyama, H. Hirai, N. Yasuda, K. Noguchi, T. Asakawa, S. Sugiyama, T. Kan, H. Kawagishi* — ■■■■-■■■■

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Das Stoffwechselprodukt 2-Aza-8-oxohypoxanthin (2) des mit Hexenringen in Verbindung stehenden 2-Azahypoxanthin (1) wurde aus Reis isoliert, was für das Vorliegen von endogenem 1 und 2 in Pflanzen spricht. 1 kann durch Umsetzung von 3 mit zunächst NaNO₂ und

anschließend NH₃ synthetisiert und danach durch Xanthin-Oxidase in 2 umgewandelt werden. Der neu entdeckte Purinpfad zu diesen Verbindungen in Reis und *Arabidopsis* folgt einer ähnlichen Route wie die chemische Synthese.