

Cesium uptake and translocation from tea cutting roots (*Camellia sinensis* L.)

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1 **Cesium uptake and translocation from tea cutting roots**
2 **(*Camellia sinensis* L.)**

3

4 **Hiroto Yamashita^{1,2}, Yoshifumi Nishina¹, Naho Komori¹, Mizuho Kamoshita¹,**
5 **Yasuhisa Oya³, Kenji Okuno³ and Akio Morita^{1,4}, Takashi Ikka^{1,4*}**

6

7 ¹Laboratory of Functional Plant Physiology, Faculty of Agriculture, Shizuoka University,
8 836 Ohya, Shizuoka, Shizuoka 422-8529, Japan

9 ²United Graduate School of Agricultural Science, Gifu University, 1-1 Yanagito, Gifu
10 501-1193, Japan

11 ³Radioscience Research Laboratory, Faculty of Science, Shizuoka University, 836 Ohya,
12 Shizuoka, Shizuoka 422-8529, Japan

13 ⁴Institute for Tea Science, Shizuoka University, 836 Ohya, Shizuoka, Shizuoka 422-8529,
14 Japan

15

16 *Corresponding author: ikka.takashi@shizuoka.ac.jp

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18

19 **Abstract**

20 To estimate the uptake of radiocesium (^{137}Cs) by tea plant roots, 1-year-old rooted tea
21 cuttings (*Camellia sinensis* L. cv. Yabukita) at the time of bud opening were cultivated
22 hydroponically for 27 days in pots containing nutrient solutions with or without $^{137}\text{CsCl}$
23 (600 Bq mL^{-1}). Total ^{137}Cs radioactivity of whole tea plants were 6.1 kBq g^{-1} dry weight.
24 The plant/solution ^{137}Cs transfer factors of different tissues were in the range of 2.6 (in
25 mature leaves) to 28.2 mL g^{-1} dry weight (in roots), which were lower than those reported
26 in wheat and spinach. In total, 69% of ^{137}Cs remained in roots and 31% was transported
27 from roots to shoots. The results indicated that ^{137}Cs was preferentially translocated to
28 new shoots, which are used for manufacturing tea, over mature leaves.

29

30 **Keywords**

31 Tea plants (*Camellia sinensis* L.); Fukushima Dai-ichi Nuclear Power Plants;
32 Radiocesium; Root uptake

33

34 **Abbreviations**

35 TFs, transfer factors; FDNPP, Japan's Fukushima Dai-ichi Nuclear Power Plant (FDNPP)

36 **1. Introduction**

37 Radioactive cesium (i.e., ^{134}Cs and ^{137}Cs) released by Tokyo Electric Power Company
38 Holdings's (TEPCO's) Fukushima Dai-ichi Nuclear Power Plant (FDNPP) accident
39 caused by the Great East Japan Earthquake and tsunami of March 11, 2011, has been
40 detected at levels that exceed the provisional permitted value in tea leaves around the east
41 area in Japan. In a report by the Ministry of Health, Labour and Welfare (2011), the first
42 crops of tea harvested in Ibaraki, Chiba, Gunma, Kanagawa and Shizuoka Prefectures
43 after the accident were contaminated with radiocesium at a higher level than the
44 provisional regulation value for raw food materials set by the Food Safety Commission
45 of Japan, that is 500 Bq kg^{-1} (sum of ^{134}Cs and ^{137}Cs activities). Therefore, as the shipment
46 of tea leaves in these areas was stopped and workers suffered significant economic
47 losses. During the 2 years following the TEPCO's FDNPP accident, Hirono and Nonaka
48 (2016) reported a exponential decrease in the concentration of ^{134}Cs and ^{137}Cs in new
49 shoots of tea plants in Shizuoka, Japan, approximately 400 km southwest from the
50 FDNPP. In a previous study, we showed that radiocesium was primarily absorbed from
51 the lower surface of tea leaves through the stomata, and that the greater part was
52 transported to newly emerged tea tissues, especially new shoots, during the new shoot
53 growth period (Ikka et al., 2018). Therefore, new shoots harvested in the first flush of
54 2011 were indirectly contaminated by FDNPP's radiocesium fallout. However, because

55 tea plants are a perennial crop, there is concern about the long-term effects of indirect
56 contamination from the soil or source organs other than the new shoots that are used to
57 make tea products (Tagami et al., 2020, 2012). To reduce radiocesium amounts in tea
58 plants, pruning was most effective measure (Hirono and Nonaka, 2016).

59 In the FDNPP accident, the soil of tea fields was contaminated by radionuclides
60 (Takeda et al., 2013). In particular, long-lived ^{137}Cs in the soil will be a source of
61 radiocesium contamination in tea plants for the long term through root uptake. In general,
62 Cs is not an essential element for plant growth and therefore its absorption mechanism
63 and use by plants has not been studied in depth. As Cs is an alkali metal like K, the two
64 have similar chemical properties and K transport systems also function in Cs uptake by
65 roots (Avery et al., 1993, 1991; Zhu and Smolders, 2000). Previous studies showed that
66 ^{137}Cs activities in the primary and secondary roots of tea plants after the Chernobyl
67 Nuclear Power Plant accident in 1986 greatly decreased in 1987, but then gradually
68 increased up to 1993 (Topcuoğlu et al., 1997). Ertel and Ziegler (1991) reported that 20%
69 of the ^{137}Cs translocated into new leaves of larch (*Larix decidua* Mill.) and about 50% of
70 that into sycamore maple (*Acer pseudoplatanus* L.) resulted from root uptake during the
71 2.5 years after the Chernobyl accident. These reports suggest that tea plants might take

72 up ^{137}Cs from contaminated soils over the long term. However, the rate of ^{137}Cs uptake
73 by tea roots has not previously been estimated.

74 The ^{137}Cs uptake activities of plants is estimated as the soil-to-plant transfer factor
75 (TF) (Ehlke and Kirchner, 2002). However, in the literature, for a number of long-lived
76 radionuclides, reported soil-to-plant TFs showed variations that exceeded three orders of
77 magnitude (Coughtrey and Thorne, 1983; Frissel, 1992). The soil-to-plant TF is a
78 macroscopic parameter that integrates various soil chemical, soil biological, hydrological,
79 physical, and plant physiological processes, each of which shows its own variability.
80 Therefore, in several plant species, TFs have been estimated in hydroponic culture by
81 determining the solution-to-plant TFs, enabling more precise measurement of the ^{137}Cs
82 uptake activities of plant roots (Smolders et al., 1997; Smolders and Shaw, 1995).

83 In this study, we examined radiocesium uptake into tea roots by applying ^{137}Cs
84 into hydroponic solution supplied to cultured tea plants, and determined the TFs of ^{137}Cs
85 in several plant parts during the development of new shoots.

86

87 **2. Materials and methods**

88 *2.1. Plant materials and hydroponic experiments*

89 One-year-old rooted tea cuttings (*Camellia sinensis* L. cv. Yabukita) were carefully
90 washed with tap water to remove soil and then transplanted to a/5,000 Wagner pots
91 supplied with continuously aerated hydroponic solution (volume 3 L) in a greenhouse in
92 Shizuoka University (Shizuoka, Shizuoka, Japan). The hydroponic solution was prepared
93 according to Konishi et al., (1985), adjusted to pH 4.2 with 1 M H₂SO₄ and renewed every
94 week until the start of treatment.

95 In 2014, we conducted the following hydroponic experiment to evaluate ¹³⁷Cs
96 uptake from tea roots. After 9 weeks, to synchronize the growth stage of tea plants, one
97 bud and three leaves of every new shoot were cut off. After 3 weeks, at the time of bud
98 opening (Supplementary Fig. 1), three tea plants were transplanted into each one new pot
99 containing the hydroponic solutions (volume 3 L) with (+Cs) or without (-Cs) using 3.7
100 MBq L⁻¹ ¹³⁷CsCl solution (Eckert and Ziegler Isotope Products, Valencia, CA, USA). At
101 the start of the treatment, the final ¹³⁷Cs concentration of the +Cs treatment solution was
102 600.0 Bq mL⁻¹. Each treatment was carried out with three replicates. During the treatment,
103 tea plants were cultivated in draft chambers of the Radiochemistry Research Institute,
104 Shizuoka University. In the draft chamber, light/dark periods were 12 h/12 h, and light
105 intensity at the canopy was kept at around 200 μmol m⁻² s⁻¹ PPFD using two white LED
106 lamps (Natural Spectrum light, VeriLux Inc., Vermont, USA). The +Cs and -Cs solutions

107 were not changed during treatment, but the solutions were maintained at a volume of 3 L
108 using ^{137}Cs hydroponic solution. After 27 d, at the opening of the third to fourth leaves, tea
109 plants were washed with tap water at least three times then wiped dry. Each tea plant (one
110 replicate) was separated into new shoots, mature leaves, upper trunk, lower trunk, and
111 roots. After measuring fresh weights, each part was kept at $-30\text{ }^{\circ}\text{C}$ until ^{137}Cs analysis.
112 Representative plants were used to perform ^{137}Cs image plate analysis.

113

114 *2.2. Measurement of ^{137}Cs radiation*

115 Collected samples were dried for 3 d at $60\text{ }^{\circ}\text{C}$, then weighed samples were placed in 10-
116 mL polystyrene tubes for radioactivity analysis. The radioactivity of ^{137}Cs was evaluated
117 by an auto-well gamma system (ARC-380CL, Aloka Inc.) calibrated using a standard
118 ^{137}Cs source with each part. Radioactivity values of tea plant samples were obtained after
119 subtraction of the background values from samples without applied ^{137}Cs .

120

121 *2.3. Calculation of ^{137}Cs transfer factors from solution to tea plants*

122 In the experiment to study ^{137}Cs uptake by tea roots, transfer factors (TFs) of ^{137}Cs from
123 the solution to tea plants (plant/solution ^{137}Cs TF; mL g^{-1} dry weight, DW) were
124 calculated as the ratio of the ^{137}Cs activity concentration in each plant part (Bq g^{-1} DW)

125 to that of the treatment solution (600 Bq mL^{-1}). ^{137}Cs radioactivity (kBq plant^{-1}) of whole
126 plants was calculated by summing the data of each plant part. And, ^{137}Cs radioactivity
127 ($\text{kBq g}^{-1} \text{ DW}$) of whole plants was calculated from the ^{137}Cs radioactivity (kBq plant^{-1})
128 and DW.

129

130 *2.4. Imaging plate analysis*

131 One set of shoots and roots of +Cs treatments was put into a polyethylene film bag, closely
132 set on the imaging plate (BAS-MS2040, FUJIFILM, Tokyo, Japan), and exposed in
133 darkness for 21 d at room temperature. Radioactivity distribution images were obtained
134 by scanning the imaging plates with the molecular measurement program FX Pro Plus
135 (BioRad Laboratories, Inc., California, USA).

136

137 *2.5. Statistical analyses*

138 Data were statistically analyzed using Tukey's honestly significant difference (HSD) test
139 to determine significant differences among groups. P -values < 0.05 were considered
140 significant. Statistical analyses were performed using R software ver. 4.0.2.

141

142 **3. Results and Discussion**

143 Each of five parts and total dry weights (DW) at the end of treatment are shown in Table
144 1. The DW of new shoots was 0.9 g plant⁻¹; two or three new shoots per plant emerged
145 during 27 d of cultivation in the draft chamber with radiocesium application, meaning
146 that it is possible to evaluate the translocation of the radiocesium from tea roots to new
147 shoots.

148 ¹³⁷Cs radioactivity and TFs from tea roots are shown in Table 1. Total ¹³⁷Cs
149 radioactivity of whole tea plants was 6.1 kBq g⁻¹ DW. Comparing the radioactivity among
150 plant parts, it was highest in roots (16.9 kBq g⁻¹ DW) and lowest in mature leaves and
151 upper trunk (1.5 and 2.3 kBq g⁻¹ DW, respectively). The radioactivity of new shoots (4.2
152 kBq g⁻¹ DW) was between that of roots and mature leaves. These results were supported
153 by the image plate analysis (Fig. 1). The results indicate that ¹³⁷Cs was preferentially
154 translocated to new shoots, which are used for manufacturing tea, rather than mature
155 leaves. The radioactivity of the lower trunk (3.2 kBq g⁻¹ DW) was tended to be higher
156 than that of the upper trunk because a part of the lower trunk was immersed in the
157 treatment solution (Supplementary Fig. 1). Thus, the ¹³⁷Cs radioactivity of the lower trunk
158 might include not only the ¹³⁷Cs transported from roots but also the ¹³⁷Cs absorbed
159 directly from the treatment solution itself and remaining adventitious roots. This means a
160 limitation of the hydroponic test using clonal tea cuttings, which have adventitious roots

161 from the base of the lower trunk. Based on ^{137}Cs radioactivity of each plant part and the
162 nutrient solution (600.0 Bq mL^{-1}), the plant/solution ^{137}Cs TFs were in the range from 2.6
163 (in mature leaves) to 28.2 mL g^{-1} (in roots) (Table 1). The ^{137}Cs TF of new shoots (6.9
164 mL g^{-1}) was around one-third of the roots' value and similar to that of the whole tea plant
165 (10.2 mL g^{-1}). In wheat (*Triticum aestivum* cv. Tonic) cultivated for 21 d in nutrient
166 solution containing 3 mM K^+ , $0\text{--}4.24 \text{ mM NH}_4^+$ or $4.24\text{--}4.98 \text{ mM NO}_3^-$, $0.25\text{--}2.49 \text{ mM}$
167 Ca^{2+} and $0.72\text{--}1.27 \text{ mM Mg}^{2+}$ with $5 \text{ Bq mL}^{-1} \text{ }^{137}\text{Cs}^+$, the plant/solution ^{137}Cs TF was 30--
168 60 mL g^{-1} for the shoot and $60\text{--}140 \text{ mL g}^{-1}$ for the roots (Smolders and Shaw, 1995). In
169 spinach (*Spinacia oleracea* L. cv. Subito), the plant/solution ^{137}Cs TF was $41\text{--}117 \text{ mL}$
170 g^{-1} , when cultivated in solution containing $5 \text{ Bq mL}^{-1} \text{ }^{137}\text{Cs}$ with the following range of
171 cationic concentrations: $0.53\text{--}10.4 \text{ mM K}$; $0\text{--}8.47 \text{ mM NH}_4$; $0.15\text{--}5.0 \text{ mM Ca}$; $0.08\text{--}2.0$
172 mM Mg (Smolders et al., 1997). It has been shown that the concentrations of the cations,
173 K^+ , NH_4^+ , Ca^{2+} , and Mg^{2+} , in the culture solution affected the ^{137}Cs uptake of plants (Zhu
174 and Smolders, 2000). The concentrations of 1 mM K^+ , 1.8 mM NH_4^+ , 0.5 mM Ca^{2+} , and
175 0.4 mM Mg^{2+} in the tea culture solution used in our experiments (from Konishi et al.,
176 1985) were within the range of those in the above-mentioned wheat and spinach culture
177 solutions. These results indicated that the ^{137}Cs uptake activity of tea plant roots from the
178 solution was lower than those of wheat and spinach, although it is necessary to consider

179 the difference in ^{137}Cs concentrations and its ratio to K of the culture solutions and the
180 ^{137}Cs adhesion on surface.

181 It is known that Cs^+ enters into plant cells through potassium ion (K^+) transporters
182 (Avery et al., 1993, 1991; Sacchi et al., 1997; Sheahan et al., 1993; Zhu and Smolders,
183 2000), therefore any factor that influenced K^+ uptake might also affect Cs^+ uptake. It was
184 reported that pH affected the activity of K^+ transporters (Maathuis and Sanders, 1996). In
185 *Riccia fluitans* cultivated in culture solution buffered at pH 6.5–9.0, Cs uptake rates under
186 K^+ deficiency showed a maximum at pH 7.5 with declines under more acid or more
187 alkaline conditions, but were not affected by pH under K^+ sufficiency (Heredia et al.,
188 2002). In our experiment, the culture solution was adjusted to an acidic condition (pH
189 4.2), because the optimum pH for tea plant growth was 4.0–5.0 in the presence of 0.4 mM
190 Al^{3+} (Konishi et al., 1985; Yamashita et al., 2020). Konishi et al. (1985) reported that 1
191 mM K^+ was adequate for tea growth in solution culture, and K uptake by tea plants was
192 stimulated under this acidic condition in the presence of Al. These results might mean
193 that the cation concentrations under the acidic conditions of our culture solutions-did not
194 bring about the lower Cs^+ uptake activity of tea plants.

195 Among five parts of tea plants, the ^{137}Cs radioactivity was the highest in roots
196 ($31.6 \text{ Bq plant}^{-1}$) followed by new shoots ($3.7 \text{ kBq plant}^{-1}$), lower trunk ($3.5 \text{ kBq plant}^{-1}$),

197 mature leaves ($3.4 \text{ kBq plant}^{-1}$), upper trunk ($3.1 \text{ kBq plant}^{-1}$) (Table 1). Thus, 69% of
198 the total amount of ^{137}Cs per plant remained in roots, and 31% was transported from roots
199 to shoots. In previous reports, tissues that contained low K concentration, such as ears,
200 fruits, or wood, were also found to be low in Cs (Zhu and Smolders, 2000). Konishi et al.
201 (1985) reported that K concentration was the highest in roots ($30.6 \text{ mg g}^{-1} \text{ DW}$), followed
202 by shoot tips ($20.3 \text{ mg g}^{-1} \text{ DW}$), which were similar to mature leaves ($18.3 \text{ mg g}^{-1} \text{ DW}$)
203 > stems ($14.8 \text{ mg g}^{-1} \text{ DW}$) in 1-year-old rooted tea cuttings cultivated in culture solution.
204 Given this report (Konishi et al., 1985) of the K distribution in tea plants, it is possible
205 that the concentration of radiocesium was also high in the roots. However, in our results,
206 the Cs distribution among the shoot tips, mature leaves, and trunk of tea plants did not
207 exactly reflect that of K (Table 1). Topcuoğlu et al. (1997) detected radiocesium only in
208 the roots of tea plants in Turkey 8 years after the Chernobyl NPP accident. This finding
209 suggested that the rate of Cs transport into the xylem in the tea plant roots was low
210 although there is a difference between hydroponics and field cultivation. It was reported
211 that an important property of plants in relation to ^{137}Cs uptake was high growth rate and
212 high biomass production in a given environment (Soudek et al., 2004). Generally, woody
213 plants showed a lower capacity for water uptake and a slower growth rate, compared with
214 annual plants. In tea plants, such properties might contribute to the low ^{137}Cs uptake.–

215

216 **4. Conclusion**

217 In conclusion, we investigated the uptake of ^{137}Cs radioactivity and its transfer
218 from tea roots to other plant parts in model hydroponic conditions. The plant/solution
219 ^{137}Cs TFs among different tissues of tea plants were in the range 2.6 (in mature leaves) to
220 28.2 mL g^{-1} (in roots), which were lower than values previously reported in wheat
221 (Smolders and Shaw, 1995) and spinach (Smolders et al., 1997). In total, 69% of ^{137}Cs
222 remained in the roots, and 31% was transported from the roots to shoots. Our results
223 indicated that ^{137}Cs was preferentially translocated to new shoots over mature leaves in
224 tea plants. Our results will contribute to understanding the long-term effects of indirect
225 radiocesium contamination from tea roots.

226

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233

234 **Conflict of interests**

235 The authors declare no conflict of interests associated with this manuscript.

236

237 **References**

238 Avery, S.V., Codd, G.A., Gadd, G.M., 1993. Transport kinetics, cation inhibition and
239 intracellular location of accumulated cesium in the green microalga *Chlorella salina*.
240 *Microbiology* 139, 827–834.

241 Avery, S.V., Codd, G.A., Gadd, G.M., 1991. Cesium accumulation and interactions with
242 other monovalent cations in the cyanobacterium *Synechocystis* PCC 6803. *J. Gen.*
243 *Microbiol.* 137, 405–413.

244 Coughtrey, P.J., Thorne, M.C., 1983. Radionuclide distribution and transport in terrestrial
245 and aquatic ecosystems. A critical review of data. Vol. 1. Balkema, Rotterdam.

246 Ehlke, S., Kirchner, G., 2002. Environmental processes affecting plant root uptake of
247 radioactive trace elements and variability of transfer factor data: a review. *J. Environ.*
248 *Radioact.* 58, 97–112.

249 Ertel, J., Ziegler, H., 1991. Cs-134/137 contamination and root uptake of different forest
250 trees before and after the Chernobyl accident. *Radiat. Environ. Biophys.* 30, 147–157.

251 Frissel, M.J., 1992. An update of the recommended soil-to-plant transfer factors of Sr-90,
252 Cs-137 and transuranics VIIIth Report of the Working Group. Soil-to-Plant Transfer
253 Factors International Union of Radioecologists. International Union of Radioecologists.

254 Heredia, M.A., Zapico, R., García-Sánchez, M.J., Fernández, J.A., 2002. Effect of
255 calcium, sodium and pH on uptake and accumulation of radiocesium by *Riccia fluitans*.
256 *Aquat. Bot.* 74, 245–256.

257 Hirono, Y., Nonaka, K., 2016. Time series changes in radiocaesium distribution in tea
258 plants (*Camellia sinensis* (L.)) after the Fukushima Dai-ichi Nuclear Power Plant accident.
259 *J. Environ. Radioact.* 152, 119–126.

260 Ikka, T., Nishina, Y., Kamoshita, M., Oya, Y., Okuno, K., 2018. Radiocesium uptake
261 through leaf surfaces of tea plants (*Camellia sinensis* L.). *J. Environ. Radioact.* 182, 70–
262 73.–

263 Konishi, S., Miyamoto, S., Taki, T., 1985. Stimulatory effects of aluminum on tea plants
264 grown under low and high phosphorus supply. *Soil Sci. Plant Nutr.* 31, 361–368.

265 Maathuis, F.J.M., Sanders, D., 1996. Mechanisms of potassium absorption by higher
266 plant roots. *Physiol. Plant.* 96, 158–168.

267 Ministry of Health, Labour and Welfare, 2011. Test Results O Radionuclide in Foods
268 since 19 March 2011. [https://www.mhlw.go.jp/stf/houdou/2r98520000029prx-](https://www.mhlw.go.jp/stf/houdou/2r98520000029prx-att/2r98520000029rvk.pdf)
269 [att/2r98520000029rvk.pdf](https://www.mhlw.go.jp/stf/houdou/2r98520000029prx-att/2r98520000029rvk.pdf) (accessed 12.21.20).

270 Smolders, E., Shaw, G., 1995. Changes in radiocaesium uptake and distribution in wheat
271 during plant development: a solution culture study. *Plant Soil* 176, 1–6.

272 Smolders, E., Sweeck, L., Merckx, R., Cremers, A., 1997. Cationic interactions in
273 radiocaesium uptake from solution by spinach. *J. Environ. Radioact.* 34, 161–170.

274 Soudek, P., Tykva, R., Vanek, T., 2004. Laboratory analyses of ¹³⁷Cs uptake by sunflower,
275 reed and poplar. *Chemosphere* 55, 1081–1087.

276 Takeda, H., Shiraki, Y., Funahashi, H., Kita, N., Yamada, Y., 2013. Vertical distributions
277 of fallout radioactive caesium in tea gardens soil in Kanagawa Prefecture. *Japanese*
278 *Journal of Soil Science and Plant Nutrition* 84, 49–52.

279 Tagami, K., Uchida, S., Ishii, N., Kagiya, S., 2012. Translocation of radiocesium from
280 stems and leaves of plants and the effect on radiocesium concentrations in newly emerged
281 plant tissues. *J. Environ. Radioact.* 111, 65–69.

282 Tagami, K., Uchida, S., Shinano, T., Pröhl, G., 2020. Comparisons of effective half-lives
283 of radiocesium in Japanese tea plants after two nuclear accidents, Chernobyl and
284 Fukushima. *J. Environ. Radioact.* 213, 106109.

285 Topcuoğlu, S., Güngör, N., Köse, A., Varinlioğlu, A., 1997. Translocation and depuration
286 of ^{137}Cs in tea plants. J. Radioanal. Nucl. Chem. 218, 263–266.

287 Yamashita, H., Fukuda, Y., Yonezawa, S., Morita, A., Ikka, T., 2020. Tissue ionome
288 response to rhizosphere pH and aluminum in tea plants (*Camellia sinensis* L.), a species
289 adapted to acidic soils. Plant-Environment Interactions 1, 152–164.

290 Zhu, Y.G., Smolders, E., 2000. Plant uptake of radiocaesium: a review of mechanisms,
291 regulation and application. J. Exp. Bot. 51, 1635–1645.

292

293 **Supplementary information**

294 **Supplementary Fig. S1 Tea plant in hydroponic culture at the time of bud opening**

295 **at the start of treatment**



296 **Table legends**

297 **Table 1 ¹³⁷Cs radioactivity and transfer factor from tea roots**

Tissues	Dry weight (g plant ⁻¹)	¹³⁷ Cs radioactivity (kBq g ⁻¹ DW)	Plant/solution ¹³⁷ Cs transfer factor (kBq plant ⁻¹)	Plant/solution ¹³⁷ Cs radioactivity	Proportion of ¹³⁷ Cs radioactivity (%)
New shoots	0.9 ± 0.5	4.2 ± 0.6	b	6.9 ± 1.1	8.2 ± 3.4
Mature leaves	2.2 ± 0.8	1.5 ± 0.4	a	2.6 ± 0.6	7.5 ± 3.1
Upper trunk	1.3 ± 0.2	2.3 ± 0.1	ab	3.9 ± 0.2	6.9 ± 0.7
Lower trunk	1.1 ± 0.3	3.2 ± 0.9	ab	5.3 ± 1.5	7.7 ± 2.5
Roots	1.9 ± 0.2	16.9 ± 1.5	c	28.2 ± 2.5	69.6 ± 12.2
Whole plants	7.5 ± 1.8	6.1 ± 0.3		10.2 ± 0.5	100.0 ± 19.4

298 Values are mean ± SD (n=3). Different letters indicate significant differences among
 299 tissues (Tukey's HSD test, *P* < 0.05).

300

301 **Figure legends**

302 **Fig. 1 Distribution of ¹³⁷Cs radioactivity after uptake and translocation by tea roots**

303 Imaging plate analysis of a 1-year-old rooted tea cutting grown for 21 d in hydroponic
304 culture with $^{137}\text{CsCl}$ (600 Bq mL^{-1}). Left and right images show the plant appearance and
305 distribution of radioactivity, respectively.

306
307

