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The natural alkaloid sanguinarine promotes the expression of heat shock protein genes in Arabidopsis

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1 Title page 23 **SHORT COMMUNICATION** 4 5 Title The natural alkaloid sanguinarine promotes the expression of heat shock protein genes in 6 7 Arabidopsis 8 9 **Authors** 10 Masakazu Hara, Ikuya Kurita Research Institute of Green Science and Technology, 11 12 Shizuoka University, 13 836 Ohya, Shizuoka 422-8529, Japan 14 15 Name and address for editorial correspondence 16 Masakazu Hara 17 Research Institute of Green Science and Technology, 18 Shizuoka University, 836 Ohya, Shizuoka 422-8529, Japan 19 20 Telephone number: +81-54-238-5134 Fax number: +81-54-238-5134 2122 E-mail address: amhara@ipc.shizuoka.ac.jp 23

#### Abstract

Small-molecule heat shock response inducers are known to enhance heat tolerance in plants. In this paper, we report that a plant alkaloid enhances the heat tolerance of *Arabidopsis*. We investigated 12 commercially available alkaloids to determine whether they enhance the heat tolerance of *Arabidopsis* seedlings by using an *in vitro* assay system with geldanamycin, which is a known heat shock response inducer, as a positive control. Accordingly we found that the isoquinoline alkaloid sanguinarine can enhance heat tolerance in *Arabidopsis*. No such effect was shown for the other 11 alkaloids. The sanguinarine treatment increased the expression of heat shock protein genes such as *HSP17.6C-CI*, *HSP70*, and *HSP90.1*, which were up-regulated by geldanamycin. Treatments with other isoquinoline alkaloids (berberine and papaverine), which showed few heat-tolerance-enhancing effects, did not promote the expression of the heat shock protein genes. These results suggest that sanguinarine influenced the heat tolerance of *Arabidopsis* by enhancing the expression of heat shock protein genes.

#### Keywords

39 Alkaloids - Arabidopsis thaliana - Geldanamycin - Heat shock protein - Heat tolerance -

40 Sanguinarine

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#### Introduction

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The extremely high temperatures caused by heat waves reduce plant production in many areas of the world (Hall 2001; Ainsworth and Ort 2010). One useful approach to suppressing the loss of plant productivity due to heat is to ameliorate the heat stress caused by high ambient temperatures. When plants are exposed to heat stress, various symptoms occur including protein denaturation, the inhibition of protein synthesis, protein degradation, increased fluidity of membrane lipids, the production of reactive oxygen species, and the suppression of photosynthesis (Wahid et al. 2007; Allakhverdiev et al. 2008; Ruelland and Zachowski 2010). In order to reduce the occurrence of such negative symptoms, plants accumulate compatible solutes, alter their hormone and growth regulator contents, enhance their antioxidative activities, and express heat shock proteins (HSPs) (Iba 2002; Kotak et al. 2007). It is believed that the expression of HSP genes is an important factor in establishing heat tolerance in plants (Huang and Xu 2008), because positive correlations between the expression levels of HSP genes and the degree of heat tolerance have been demonstrated (Wang et al. 2004; Wahid et al. 2007; Waters 2013; Zhang et al. 2014). Controlling the HSP expression can be an effective technique to ameliorate the heat damage sustained by plants.

Some low-molecular-weight compounds have shown an ability to enhance heat tolerance in plants. The application of salicylic acid (SA) (Dat et al. 1998; Senaratna et al. 2000), glycinebetaine (GB) (Sakamoto and Murata 2002), polyamines (Wahid et al. 2007), geldanamycin (GDA) (Yamada et al. 2007), and isothiocyanates (ITCs) (Hara et al. 2013) has been shown to enhance the heat tolerance of plants at the laboratory level. Among the known enhancers, GDA has been well characterized in terms of the mechanism by which it enhances heat tolerance. GDA was originally found in *Streptomyces hygroscopicus* as an antibiotic. Since it was demonstrated that GDA shows antitumor activities by inhibiting the HSP known as HSP90s in mammals (Trepel et al. 2010; Roe et al. 1999), GDA came to be used as an HSP90 inhibitor. Active HSP90 negatively regulates the action of heat shock factors (HSFs) by binding them. HSP90 inhibitors like GDA activate HSFs by inactivating HSP90, resulting in the induction of a strong heat shock response. It was proposed that the increase in the heat shock response induced by GDA enhances the heat tolerance of plants (Yamada et al. 2007; Yamada and Nishimura 2008). This suggests that other compounds showing GDA-like activities may enhance the heat tolerance of plants.

GDA is an N-containing macrocyclic polyketide showing antimicrobial activity. This suggests that natural N-containing antimicrobial products such as plant alkaloids may show heat-tolerance-enhancing effects. The alkaloids are one of the most diverse groups of secondary metabolites found mainly in plants (Roberts and Wink 1998). Among them berberine, cocaine, colchicine, mescaline, quinine, and reserpine are examples of antimicrobial alkaloids (Cowan 1999). In order to find alkaloids that show heat-tolerance-enhancing effects, 12 commercially available alkaloids were screened. Accordingly we found that the isoquinoline alkaloid sanguinarine showed a heat-tolerance-enhancing effect.

## **Materials and methods**

Chemicals

- Berberine, sanguinarine, reserpine, and trigonelline were purchased from Sigma (Tokyo, Japan).
- 90 Emetine, noscapine, papaverine, yohimbine, caffeine, quinine, capsaicin, and colchicine were
- 91 obtained from Wako (Tokyo, Japan). GDA was obtained from LC Laboratories (MA, USA).

93 Heat-tolerance screening

Solidified culture medium (1/5 MS medium with 0.8% agar and 1% sucrose, pH 5.8) was

prepared in 6-well plates (TPP, Trasadingen, Switzerland, #92406) under sterile conditions. A sterile filter paper disk (No. 1, Advantec, Tokyo, Japan) was placed on the surface of the solid medium in each well. Surface-sterilized seeds of Arabidopsis thaliana (L.) Heynh. (ecotype Columbia) were sown onto the filter paper at a rate of 10 seeds per well. After vernalization for 2 days at 6°C, the plates were placed in the growth chamber (NK System, Tokyo, Japan) for 6 days under the following conditions: a 16-h day (60 μmol m<sup>-2</sup> s<sup>-1</sup>)/8-h night cycle at 22°C. At this step, the seedlings were thinned out to 6 per well. The seedlings with the filter paper were inoculated onto the test medium in the 6-well plates 6 days after germination (DAG). Test compounds were freshly dissolved in dimethyl sulfoxide (DMSO). The concentrations of compounds other than sanguinarine were 0 (control), 0.25, 2.5, 25, and 250 mM. The sanguinarine concentrations were 0 (control), 0.025, 0.25, 2.5, and 25 mM. The test medium (4 ml in each well) was prepared by combining the test DMSO solution (8 µl) and the 1/5 MS medium with 0.8% agar and 1% sucrose (3992 µl). The pH of the test medium was checked using pH indicator strips (Merck, Tokyo, Japan). The plates were incubated under the same conditions as given above for 24 h, and then at 7 DAG the plants were heated as follows. The plates were covered with a polyethylene zipper bag (4-F, 120 nm x 170 mm, System Polymer, Tokyo, Japan) and were totally soaked at 47°C for 1 h. Control plates were placed in the same growth chamber for 1 h. After the plates containing heat-treated and control plants were maintained under the same cultivation conditions for 3 days, the fresh weights of the plants were measured (10 DAG).

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## Transcripts analysis

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119 Transcripts accumulation of the HSP17.6C-CI (At1g53540), HSP70 (At3g12580), HSP90.1 120 (At5g52640), and ACTIN-2 (At3g18780) genes in Arabidopsis was analyzed by reverse 121 transcription-polymerase chain reaction (RT-PCR) according to the method given in a previous 122 report (Hara et al. 2013). Arabidopsis seedlings were treated with sanguinarine, GDA, berberine, 123 and papaverine at the concentration of 50 µM in the 6-well plates as described above. The 6 DAG seedlings with the filter paper were inoculated onto the medium containing alkaloids, and 124 125 the seedlings were harvested at 0, 1, 6, and 24 h after the inoculation. To provide the heat shock, 126 the seedlings were incubated at 37°C. The harvesting periods were the same as in the alkaloid 127 treatments. Total RNA was extracted from the whole seedlings with the RNeasy Plant Mini Kit 128 (Qiagen, Tokyo, Japan). Total RNA (0.5 μg) was subjected to reverse transcription (45°C for 30 129 min), and then underwent PCR with cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 60 s 130 (24 to 40 cycles were performed for HSP17.6C-CI, 18 to 30 cycles for HSP70, 18 to 32 cycles 131 for HSP90.1, and 18 to 24 cycles for ACTIN-2). The primers were

| 132 | 5'-CTCTAATTCCAAGCATC-3' (sense) and 5'-CCAGAGATATCAATGGAC-3' (antisense) for                  |
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| 133 | HSP17.6C-CI, 5'-GATCTCTAATAATGGCGGG-3' (sense) and  |
| 134 | 5'-CCAGTTTCAGAGTGACATAG-3' (antisense) for <i>HSP70</i> ,                                     |
| 135 | 5'-CATCAACACGTTCTACAGC-3' (sense) and 5'-GTTCCAGACCTCGCAATGG-3'                               |
| 136 | (antisense) for HSP90.1, and 5'-ACCTTGCTGGACGTGACCTTACTGAT-3' (sense) and                     |
| 137 | 5'-GTTGTCTCGTGGATTCCAGCAGCTT-3' (antisense) for ACTIN-2. The amplified products               |
| 138 | were analyzed by 1% agarose gel electrophoresis. The band intensity was measured using NIH    |
| 139 | Image software (http://rsbweb.nih.gov/nih-image/) to determine the cycle where the PCR        |
| 140 | product was logarithmically amplified. The relative amounts of transcripts were calculated by |
| 141 | standardizing the band intensities at the zero time point in each graph.                      |
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Statistical analysis

Data for *P* values were analyzed by Student's *t* test at a significance level of 0.05.

### **Results and discussion**

Heat-tolerance screening

- We listed the plant alkaloids that showed antimicrobial activities. Although there are a large number of antimicrobial alkaloids, we randomly chose commercially available ones from each major subgroup of natural alkaloids. Berberine and sanguinarine were the isoquinoline alkaloids, reserpine and yohimbine were the indole alkaloids, caffeine was the purine alkaloid, quinine was the quinoline alkaloid, and trigonelline was the pyridine alkaloid. These alkaloids have been reported to have antimicrobial activities (Cowan 1999; Godowski 1989; Almeida et al. 2006; Ozçelik et al. 2011). Two antimicrobial alkaloids which do not belong to the major subgroups (capsaicin and colchicine) were also chosen (Jones et al. 1997; Cowan 1999). Because sanguinarine was found during the screening to show heat-tolerance-enhancing activity, we added other isoquinoline alkaloids including emetine, noscapine, and papaverine to the test list. Table 1 shows the 12 alkaloids and GDA as a positive control with their typical activities. Their structures are shown in Supplemental Fig. 1.
- We designed a heat-tolerance screening system using sterile *Arabidopsis* seedlings. The optimum heat shock condition (at 47°C for 1 h) was determined by changing the incubation temperature from 45 to 50°C. We first tested GDA, the positive control for the heat-tolerance-enhancing effect, in order to validate the efficiency of the heat-tolerance

screening (Fig. 1a). When non-GDA-treated *Arabidopsis* seedlings were not exposed to heat, the fresh weights of the seedlings increased from 7 DAG (preHS) to 10 DAG (0 µM, white bar). However, the fresh weights of *Arabidopsis* seedlings subjected to heat shock at 10 DAG (0 µM, black bar) were lower than the fresh weights of the seedlings at 7 DAG, indicating that the seedlings were damaged by the heat shock. The addition of GDA at 50 µM ameliorated the damage to the seedlings (50 µM, black bar). This GDA concentration was the same as the effective concentration described in a previous report (Yamada et al., 2007). This suggests that our screening system worked as expected.

First, nine antimicrobial alkaloids, i.e., berberine, sanguinarine, reserpine, yohimbine, caffeine, quinine, trigonelline, capsaicin, and colchicine, were investigated in the heat-tolerance screening. Except for sanguinarine, their concentrations were 0, 0.5, 5, 50, and 500  $\mu$ M. The sanguinarine concentrations were 0, 0.05, 0.5, 5, and 50  $\mu$ M. As a result, we found that only sanguinarine showed heat-tolerance-enhancing effect among the alkaloids; another isoquinoline alkaloid, berberine, did not show such activity. We subjected three additional isoquinoline alkaloids (emetine, noscapine, and papaverine) to the screening and found that out of the five isoquinoline alkaloids tested, only sanguinarine showed heat-tolerance-enhancing activity (Table 1).

The dose-dependency of the heat-tolerance-enhancing effect of sanguinarine is shown in Fig. 1b. The fresh weight of 5  $\mu$ M sanguinarine-treated seedlings was significantly higher than that of non-treated seedlings after exposure to heat (Fig. 1b, black bars). On the other hand, treatment with berberine and papaverine did not alter the fresh weights of seedlings after the heat shock (Figs. 1c, d). Similar negative results were obtained when nine alkaloids including emetine, noscapine, yohimbine, reserpine, caffeine, quinine, trigonelline, capsaicin, and colchicine were tested.

Expression of heat shock protein genes by sanguinarine

It is known that the expression of HSP genes is related to heat tolerance in plants (Wang et al. 2004; Wahid et al. 2007; Zhang et al. 2014). GDA, which is a heat-tolerance enhancer, promotes some HSP genes in *Arabidopsis*. *HSP17.6C-CI*, *HSP70*, and *HSP90.1* are reported to be major GDA-inducible HSP genes (Yamada et al. 2007; Yoshida et al. 2011). This suggests that sanguinarine may also promote the expression of these three genes. Transcript accumulations of *HSP17.6C-CI*, *HSP70*, and *HSP90.1* in plants treated with sanguinarine, GDA, berberine, papaverine, and heat were analyzed. The *Actin-2* gene was tested as a housekeeping gene. The administration of sanguinarine, GDA, and heat promoted the expression of *HSP17.6C-CI*, *HSP70*, and *HSP90.1* (Figs. 2a, b, e). However, treatment with berberine and papaverine did not

change the transcript levels of *HSP17.6C-CI*, *HSP70*, or *HSP90.1* (Figs. 2c, d). *Actin-2* transcripts were not affected in any of the cases. These results indicate that sanguinarine promoted the expression of the HSP genes that were up-regulated by GDA and heat.

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Proposed mechanism of sanguinarine

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We searched for heat-tolerance enhancers among 12 well-known alkaloids that have highly diverse structures (Supplemental Fig. 1). Through the screening, we found that sanguinarine enhanced the heat tolerance of *Arabidopsis*. Sanguinarine, which is an alkaloid produced by Papaveraceae plants such as *Sanguinaria canadensis*, *Papaver somniferum*, *Bocconia frutescens*, *Chelidonium majus*, and *Macleya cordata*, has been studied for possible medicinal applications. As a result, sanguinarine has been found to have diverse effects such as antihypertensive, antiplatelet, antimicrobial, and anticarcinogenic activities (Mackraj et al. 2008). However, there

has been no report describing the physiological action of sanguinarine in plants.

Sanguinarine is the first true alkaloid which shows a heat-tolerance-enhancing effect in plants. Although several compounds, such as GDA (Yamada et al. 2007), benzyl alcohol (Saidi et al. 2005), salicylate, chlorophenols, celastrol (Saidi et al. 2007), and ITC (Hara et al. 2013) have been previously demonstrated to be heat-shock-response enhancers, GDA is the only compound whose target molecule, i.e., HSP90, has been determined. It was reported that part of the heat shock response is regulated by HSP90 (Yamada and Nishimura, 2008). Under normal conditions, HSF is inactivated by HSP90. GDA, after it is taken up into plant cells, may inhibit HSP90. This inhibition promotes the release of active HSF, and then the HSP genes are expressed. Sanguinarine enhanced the expressions of the three HSP genes (HSP17.6C-CI, HSP70, and HSP90.1) that were up-regulated by GDA. Interestingly, the time-course patterns of the gene expressions were similar between GDA and sanguinarine (Figs. 2a, b). These results suggest that sanguinarine might promote the heat shock response of Arabidopsis by a mechanism similar to that of GDA, although little structural commonality was found between the two compounds. Berberine, which possesses a benzylisoquinoline structure, did not promote the heat shock response. This difference in their ability to promote the heat shock response of Arabidopsis may reveal structural differences between benzylisoquinoline (berberine) and benzophenanthridine (sanguinarine). Although the structural formula of berberine is similar to that of sanguinarine (Supplemental Fig. 1), berberine and sanguinarine shows buckled and planar structures, respectively (Maiti and Kumar 2010). The difference in spatial structures may

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Possible application of sanguinarine

affect the activities for promoting the heat shock response.

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| 241 | In present agricultural practice, heat-tolerance breeding is a major method of maintaining crop                   |
| 242 | productivity under extremely high temperatures (Wahid et al., 2007). However, efficient                           |
| 243 | breeding methods remain to be established in many species. Alternative methods such as the                        |
| 244 | application of heat-tolerance enhancers should be considered. Sanguinarine may be effective as                    |
| 245 | a heat-tolerance enhancer. Pharmacological and toxicological studies have suggested that                          |
| 246 | rigorous toxicity testing is needed before the application of this compound can be put into                       |
| 247 | practice (Mackraj et al. 2008). Recently, sanguinarine has been applied to livestock production                   |
| 248 | by expecting better performance (Kosina et al. 2004; Vieira et al. 2008; Yao et al. 2010).                        |
| 249 | Sanguinarine may be found to be effective as a heat-tolerance enhancer in the fields of                           |
| 250 | agriculture and environmental protection.   |
| 251 |   |
| 252 | <b>Author contributions</b> MH and IK planned the research, performed the experiments, analyzed                   |
| 253 | the data, and approved the manuscript.  |
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| 259 | Organization.   |
| 260 |   |
| 261 | References  |
| 262 |   |
| 263 | Ainsworth EA, Ort DR (2010) How do we improve crop production in a warming world? Plant                           |
| 264 | Physiol 154:526-530   |
| 265 |   |
| 266 | Allakhverdiev SI, Kreslavski VD, Klimov VV, Los DA, Carpentier R, Mohanty P (2008) Heat                           |
| 267 | stress: an overview of molecular responses in photosynthesis. Photosynth Res 98:541-550                           |
| 268 |   |
| 269 | Almeida AA, Farah A, Silva DA, Nunan EA, Glória MB (2006) Antibacterial activity of coffee                        |
| 270 | extracts and selected coffee chemical compounds against enterobacteria. J Agric Food Chem                         |
| 271 | 54:8738-8743  |
| 272 |   |
| 273 | Cowan MM (1999) Plant products as antimicrobial agents. Clin Microbiol Rev 12:564-582                             |
| 274 |   |
| 275 | Dat JF, Lopez-Delgado H, Foyer CH, Scott IM (1998) Parallel changes in H <sub>2</sub> O <sub>2</sub> and catalase |

| 276        | during thermotolerance induced by salicylic acid or heat acclimation in mustard seedlings. Plant   |
|------------|--|
| 277        | Physiol 116:1351-1357  |
| 278        |  |
| 279<br>280 | Godowski KC (1989) Antimicrobial action of sanguinarine. J Clin Dent 1:96-101  |
| 281        | Hall AE (2001) Crop Responses to Environment. CRC Press LLC, Boca Raton, Florida   |
| 282        | W. M. W. and T. L. W. (2010). A latent of the first of the state of th |
| 283        | Hara M, Harazaki A, Tabata K (2013) Administration of isothiocyanates enhances heat tolerance  |
| 284        | in Arabidopsis thaliana. Plant Growth Regul 69:71-77   |
| 285        |  |
| 286        | Huang B, Xu C (2008) Identification and characterization of proteins associated with plant   |
| 287        | tolerance to heat stress. J Integr Plant Biol 50:1230-1237   |
| 288        |  |
| 289        | Iba K (2002) Acclimative response to temperature stress in higher plants: approaches of gene   |
| 290        | engineering for temperature tolerance. Annu Rev Plant Biol 53:225-245  |
| 291        |  |
| 292        | Jones NL, Shabib S, Sherman PM (1997) Capsaicin as an inhibitor of the growth of the gastric   |
| 293        | pathogen Helicobacter pylori. FEMS Microbiol Lett 146:223-227  |
| 294        |  |
| 295        | Kosina P, Walterová D, Ulrichová J, Lichnovský V, Stiborová M, Rýdlová H, Vicar J, Krecman   |
| 296        | V, Brabec MJ, Simánek V (2004) Sanguinarine and chelerythrine: assessment of safety on pigs  |
| 297        | in ninety days feeding experiment. Food Chem Toxicol 42:85-91  |
| 298        |  |
| 299        | Kotak S, Larkindale J, Lee U, von Koskull-Döring P, Vierling E, Scharf KD (2007) Complexity  |
| 300        | of the heat stress response in plants. Curr Opin Plant Biol 10:310-316   |
| 301        |  |
| 302        | Mackraj I, Govender T, Gathiram P (2008) Sanguinarine. Cardiovasc Ther 26:75-83  |
| 303        |  |
| 304        | Maiti M, Kumar GS (2010) Polymorphic nucleic acid binding of bioactive isoquinoline  |
| 305        | alkaloids and their role in cancer. J Nucleic Acids (doi: 10.4061/2010/593408)   |
| 306        |  |
| 307        | Ozçelik B, Kartal M, Orhan I (2011) Cytotoxicity, antiviral and antimicrobial activities of  |
| 308        | alkaloids, flavonoids, and phenolic acids. Pharm Biol 49:396-402   |
| 309        |  |
| 310        | Roberts MF, Wink M (1998) Alkaloids: biochemistry, ecology, and medicinal applications.  |

311

Plenum Press, New York

312 313 Roe SM, Prodromou C, O'Brien R, Ladbury JE, Piper PW, Pearl LH (1999) Structural basis for 314 inhibition of the Hsp90 molecular chaperone by the antitumor antibiotics radicicol and 315 geldanamycin. J Med Chem 42:260-266 316 317 Ruelland E, Zachowski A (2010) How plants sense temperature. Environ Exp Bot 69:225-232 318 319 Saidi Y, Domini M, Choy F, Zryd JP, Schwitzguebel JP, Goloubinoff P (2007) Activation of the 320 heat shock response in plants by chlorophenols: transgenic *Physcomitrella patens* as a sensitive 321 biosensor for organic pollutants. Plant Cell Environ 30:753-763 322323 Saidi Y, Finka A, Chakhporanian M, Zrÿd JP, Schaefer DG, Goloubinoff P (2005) Controlled 324expression of recombinant proteins in Physcomitrella patens by a conditional heat-shock 325 promoter: a tool for plant research and biotechnology. Plant Mol Biol 59:697-711 326 327 Sakamoto A, Murata N (2002) The role of glycine betaine in the protection of plants from 328 stress: clues from transgenic plants. Plant Cell Environ 25:163-171 329 330 Senaratna T, Touchell D, Bunn E, Dixon K (2000) Acetyl salicylic acid (Aspirin) and salicylic 331 acid induce multiple stress tolerance in bean and tomato plants. Plant Growth Regul 30:157-161 332 333 Trepel J, Mollapour M, Giaccone G, Neckers L (2010) Targeting the dynamic HSP90 complex 334 in cancer. Nat Rev Cancer 10:537-549 335 336 Vieira SL, Oyarzabal OA, Freitas DM, Berres J, Pena JEM, Torres CA, Coneglian JLB (2008) 337 Performance of broilers fed diets supplemented with sanguinarine-like alkaloids and organic 338 acid. J Appl Poult Res 17:128–133 339 340 Wahid A, Gelani S, Ashraf M, Foolad MR (2007) Heat tolerance in plants: An overview. 341 Environ Exp Bot 61:199-223 342 343 Wang W, Vinocur B, Shoseyov O, Altman A (2004) Role of plant heat-shock proteins and 344 molecular chaperones in the abiotic stress response. Trends Plant Sci 9:244-252 345 346 Waters ER (2013) The evolution, function, structure, and expression of the plant sHSPs. J Exp 347 Bot 64:391-403

| 348 |   |
|-----|---|
| 349 | Yamada K, Fukao Y, Hayashi M, Fukazawa M, Suzuki I, Nishimura M (2007) Cytosolic HSP90          |
| 350 | regulates the heat shock response that is responsible for heat acclimation in Arabidopsis       |
| 351 | thaliana. J Biol Chem 282:37794-37804   |
| 352 |   |
| 353 | Yamada K, Nishimura M (2008) Cytosolic heat shock protein 90 regulates heat shock               |
| 354 | transcription factor in Arabidopsis thaliana. Plant Signal Behav 3:660-662                      |
| 355 |   |
| 356 | Yao JY, Shen JY, Li XL, Xu Y, Hao GJ, Pan XY, Wang GX, Yin WL (2010) Effect of                  |
| 357 | sanguinarine from the leaves of Macleaya cordata against Ichthyophthirius multifiliis in grass  |
| 358 | carp (Ctenopharyngodon idella). Parasitol Res 107:1035-1042                                     |
| 359 |   |
| 360 | Yoshida T, Ohama N, Nakajima J, Kidokoro S, Mizoi J, Nakashima K, Maruyama K, Kim JM,           |
| 361 | Seki M, Todaka D, Osakabe Y, Sakuma Y, Schöffl F, Shinozaki K, Yamaguchi-Shinozaki K            |
| 362 | (2011) Arabidopsis HsfA1 transcription factors function as the main positive regulators in heat |
| 363 | shock-responsive gene expression. Mol Genet Genomics 286:321-332                                |
| 364 |   |
| 365 | Zhang L, Zhang Q, GaoY, Pan H, Shi S, Wang Y (2014) Overexpression of heat shock protein        |
| 366 | gene PfHSP21.4 in Arabidopsis thaliana enhances heat tolerance. Acta Physiol Plant              |
| 367 | 36:1555-1564  |
| 368 |   |
| 369 | Conflict of Interest Statement  |
| 370 | The authors declare that they have no conflict of interest.                                     |
| 371 |   |

Table 1. Effects of natural alkaloids on heat tolerance of Arabidopsis thaliana

| Classes and names      | Typical activity             | Heat tolerance enhancement (this study)* |
|------------------------|------------------------------|--|
| Isoquinoline alkaloids |                              |  |
| Berberine              | Antimicrobial, carcinostatic | No                                       |
| Emetine                | Antiprotozoal                | No                                       |
| Noscapine              | Antitussive                  | No                                       |
| Papaverine             | Antispasmodic                | No                                       |
| Sanguinarine           | Antimicrobial, carcinostatic | Yes                                      |
| Indole alkaloids       |                              |  |
| Reserpine              | Antimicrobial, antipsychotic | No                                       |
| Yohimbine              | Antimicrobial, stimulant     | No                                       |
| Purine alkaloid        |                              |  |
| Caffeine               | Antimicrobial, stimulant     | No                                       |
| Quinoline alkaloid     |                              |  |
| Quinine                | Antimalarial, antimicrobial  | No                                       |
| Pyridine alkaloid      |                              |  |
| Trigonelline           | Antimicrobial, hypoglycemic  | No                                       |
| Others                 |                              |  |
| Capsaicin              | Antimicrobial, irritant      | No                                       |
| Colchicine             | Antigout, antimicrobial      | No                                       |
| Positive control       |                              |  |
| Geldanamycin           | Antimicrobial, carcinostatic | Yes                                      |

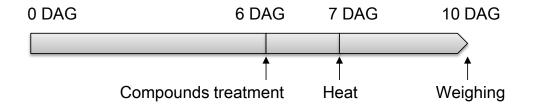
<sup>\*</sup>Positive (Yes) and negative (No) judgments of the heat-tolerance-enhancing effects were made on the basis of significant differences between the fresh weights of control plants and those of compound-treated plants 3 days after heat shock.

Test concentrations were 0.05, 0.5, 5, and 50  $\mu M$  (sanguinarine) and 0.5, 5, 50, and 500  $\mu M$  (other alkaloids).

## Figure legends

**Fig. 1.** Effects of pre-treatment with the test compounds on the growth of *Arabidopsis* seedlings after heat shock. The scheme of the experimental schedule is shown at the top. The fresh weights of the seedlings are shown. Geldanamycin (a), sanguinarine (b), berberine (c), and papaverine (d) were tested. The fresh weights of the seedlings just before the heat shock at 7 DAG are labeled preHS. White and black bars represent plants not exposed to heat and plants exposed to heat, respectively. Values and bars represent means  $\pm$  SD (n=12). Asterisks show significant differences (p < 0.05) as determined by Student's t-test in a comparison between no treatment (0  $\mu$ M) and the compound treatments.

**Fig. 2.** Effects of test compounds and heat on the expression of *HSP* genes in *Arabidopsis* seedlings. Sanguinarine (a), geldanamycin (b), berberine (c), and papaverine (d) were tested. Heat (e) was a positive control. Four genes (*HSP17.6C-CI*, *HSP70*, *HSP90.1*, and *ACTIN-2*) were analyzed. The relative accumulation of transcripts was determined by RT-PCR. Values of the 0 h controls were standardized. Values and bars represent means  $\pm$  SD (n=3). Asterisks show significant differences (p < 0.05) as determined by Student's *t*-test in a comparison between no incubation (0 h) and incubation.



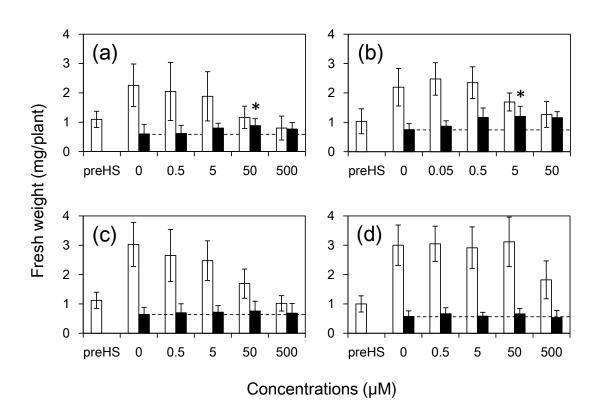


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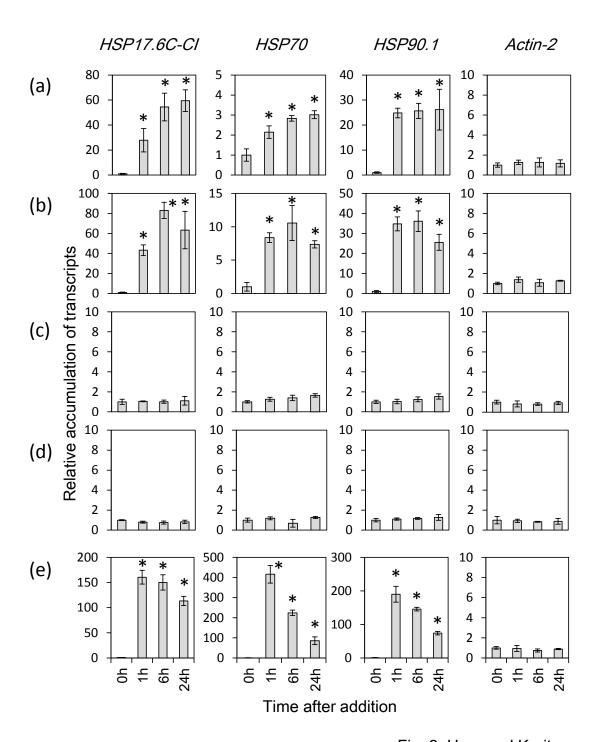
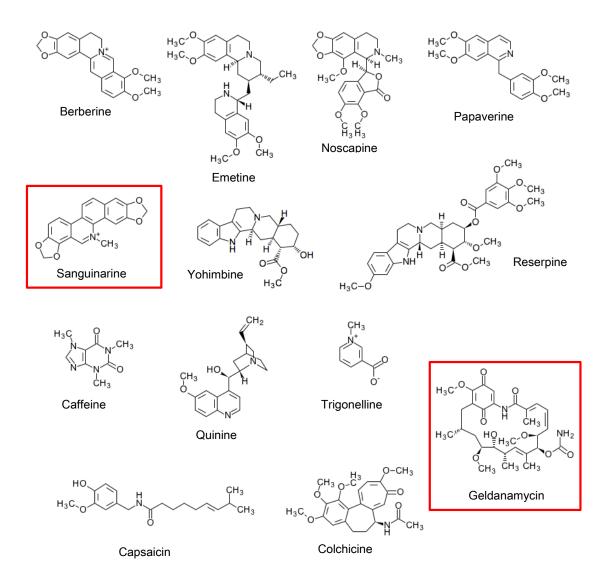


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**Supplemental Fig. 1.** Structures of alkaloids and geldanamycin tested in this study. Data were obtained from the web site of KEGG (http://www.genome.jp/kegg/kegg2.html). Sanguinarine and geldanamycin, which showed heat-tolerance-enhancing effects in our experiments, are highlighted by red frames. **Title**: The natural alkaloid sanguinarine promotes the expression of heat shock protein genes in *Arabidopsis* 

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