

## A new compound from the mushroom *Tricholoma flavovirens*

著者	Qiu Weitao, Kobori Hajime, Suzuki Tomohiro, Choi Jae-Hoon, Deo Vipin Kumar, Hirai Hirofumi, Kawagishi Hirokazu
journal or publication title	Bioscience, Biotechnology, and Biochemistry
volume	78
number	5
page range	755-757
year	2014-05-15
出版者	Taylor & Francis
権利	This is an electronic version of an article published in Bioscience, Biotechnology, and Biochemistry Volume 78, Issue 5, pages 755-757, 2014. Bioscience, Biotechnology, and Biochemistry is available online at: <a href="http://www.tandfonline.com/Article">www.tandfonline.com/Article</a> DOI; 10.1080/09168451.2014.905174.
URL	<a href="http://hdl.handle.net/10297/8617">http://hdl.handle.net/10297/8617</a>

doi: 10.1080/09168451.2014.905174

1 Running title: A New Compound from *Tricholoma flavovirens*

2

3 **A New Compound from the Mushroom *Tricholoma flavovirens***

4

5 Weitao QIU,<sup>1</sup> Hajime KOBORI,<sup>2</sup> Tomohiro SUZUKI,<sup>3</sup> Jae-Hoon CHOI,<sup>3</sup> Vipin Kumar

6 DEO,<sup>3</sup> Hirofumi HIRAI,<sup>1,3</sup> and Hirokazu KAWAGISHI<sup>1,2,3</sup>†

7

8 <sup>1</sup> *Graduate School of Agriculture, Shizuoka University, 836 Ohya, Suruga-ku, Shizuoka*  
9 *422-8529, Japan*

10 <sup>2</sup> *Graduate School of Science and Technology, Shizuoka University, 836 Ohya,*  
11 *Suruga-ku, Shizuoka 422-8529, Japan*

12 <sup>3</sup> *Research Institute of Green Science and Technology, Shizuoka University, 836 Ohya,*  
13 *Suruga-ku, Shizuoka 422-8529, Japan*

14

15 Received September 30, 2013; Accepted December 26, 2013

16

17 † To whom all correspondence should be addressed. Tel/Fax: +81-54-238-4885;

18 E-mail: achkawa@ipc.shizuoka.ac.jp

19

20

21

22

23

24

25

26

27

28

1 A novel compound (**1**) and a known one (**2**) were isolated from the fruiting  
2 bodies of *Tricholoma flavovirens*. Their structures were determined by the  
3 interpretation of spectroscopic data. Both compounds showed inhibition effects on the  
4 growth of hypocotyl of lettuce with significant differences. In addition, compound **1**  
5 showed a promotion effect on the growth of root with significant differences and **2** had  
6 the similar tendency to promote the growth.

7  
8 **Key words:** mushroom; *Tricholoma flavovirens*; structural determination; plant growth  
9 activity

10  
11 A well known axiom is that “plants act as producers, animals act as consumers,  
12 and fungi act as restorers and decomposers”. Fungi, including mushroom, play an  
13 important role in ecological balance as it can restore the nutrients used by plants and  
14 animals back to the land. We are interested in biological activity of components from  
15 mushroom towards plants and have reported isolation of some compounds that regulate  
16 lettuce growth.<sup>1-3)</sup> Using the assay evaluating growth-regulating activity toward lettuce,  
17 we screened extracts of various mushrooms and found relatively strong inhibitory  
18 activity in hexane soluble part of the extracts of the mushroom *Tricholoma flavovirens*.

19 Since ancient times *T. flavovirens* (English name, yellow knight; Japanese name,  
20 kishimeji) belonging to the family Tricholomataceae is known as an eatable mushroom  
21 throughout the world. Till now, few chemical studies were, there, so with the purpose  
22 to find novel constituents with activity from *T. flavovirens*, we started this study.

23 Here we describe the isolation, structural determination of a novel compound  
24 and a known one from the fruiting bodies of the fungus along with the biological  
25 activity of the compounds.

26 Fresh fruiting bodies of *T. flavovirens* were extracted with EtOH and then with  
27 acetone. After the solutions were combined and concentrated, they were partitioned  
28 between hexane and H<sub>2</sub>O, EtOAc and H<sub>2</sub>O, and then *n*-BuOH and H<sub>2</sub>O. The

1 hexane-soluble part was fractionated by repeated chromatography. As a consequence,  
2 two compounds (**1** and **2**) were purified.

3 Compound **1** isolated as yellow oil with a molecular formula determined as  
4  $C_{12}H_{15}NO$  by HRESIMS at  $m/z$  188.1053  $[M - H]^-$  (calcd. for  $C_{12}H_{14}NO$  188.1075),  
5 indicating presence of six degrees of unsaturation in the molecule. The structure of **1**  
6 was elucidated by interpretation of NMR spectra including DEPT, COSY, HMQC, and  
7 HMBC (Fig. 1) with the complete assignment of protons and carbons of NMR was  
8 accomplished as shown in Table 1. The DEPT experiment indicated the presence of  
9 two methyls, two methylenes, four methines and four quaternary carbons. The  
10 structure of 2-methylindole was elucidated by the HMBC correlations (H-1/C-2, C-3,  
11 C-3a, C-7a; H-3/C-2, C-3a, C-7a; H-5/C-3a, C-7; H-6/C-4, C-7a; H-7/C-3a, C-5;  
12 H-8/C-2, C-3) and the COSY correlations (H-5/H-6, H-6/H-7). The HMBC  
13 correlations (H-1'/C-2'; H-2'/C-1', C-3'; H-3'/C-2') and the COSY correlations  
14 (H-1'/H-2') indicated the presence of the ethoxymethyl group. The connection  
15 between the 2-methylindole and ethoxymethyl moiety was confirmed by the HMBC  
16 correlations (H-3'/C-3a, C-4, C-5). As a result, the structure **1** was determined as  
17 shown.

Fig. 1

Table 1

18 Compound **2** was identified as 4-methoxymethyl-2-methylindole and has been  
19 isolated from the fruiting bodies of *Tricholoma sciodes* and *Tricholoma virgatum*.<sup>4)</sup>  
20 However, no known biological activity of the compound has yet been reported.

21 Both compounds **1** and **2** showed inhibition effects on the growth of hypocotyl  
22 of lettuce at 1  $\mu\text{mol}$ /paper with significant differences. In addition, compound **1**  
23 showed a promotion effect on the growth of root at  $10^{-1}$   $\mu\text{mol}$ /paper with significant  
24 differences and **2** had the similar tendency to promote the growth (Fig. 2). The result  
25 indicated that compounds **1** and **2** possessed similar growth regulation activity against  
26 lettuce.

Fig. 2

27  
28

## 1 *Experimental*

2 *General experiments.* <sup>1</sup>H-NMR spectra (one- and two-dimensional) were recorded  
3 on a Jeol lambda-500 spectrometer at 500 MHz, while <sup>13</sup>C-NMR spectra were recorded  
4 by the same instrument at 125 MHz. A JASCO grating infrared spectrophotometer was  
5 used to record the IR spectra. The HRESIMS data were measured by a JMS-T100LC  
6 mass spectrometer. HPLC separation was performed with a Jasco Gulliver system  
7 using a reverse-phase HPLC column (Cosmosil  $\pi$ NAP Waters, Nacalai tesque, Japan).  
8 Silica gel plate (Merck F<sub>254</sub>), silica gel 60N (Merck 100-200 mesh), and C<sub>18</sub>-OPN  
9 (Cosmosil 140  $\mu$ m) were used for analytical TLC and for flash column  
10 chromatography, respectively.

11 *Fungal strain and plant materials.* Mature fruiting bodies of *T. flavovirens*  
12 were collected at Narusawa village, Yamanashi Prefecture in Japan. Lettuce seeds  
13 (*Lactuca sativa* L. cv. Great Lakes 366; Takii Co., Ltd., Japan) were used in this study.

14 *Extraction and isolation.* The fresh fruiting bodies of *T. flavovirens* (20.6 kg)  
15 were extracted with EtOH (42 L, 3 times) and then with acetone (20 L, 3 times). After  
16 the solutions were combined and concentrated under reduced pressure, the concentrate  
17 was partitioned between hexane and H<sub>2</sub>O, EtOAc and H<sub>2</sub>O, and then *n*-BuOH and H<sub>2</sub>O.  
18 The hexane-soluble part (39.8 g) was fractionated by silica gel flash column  
19 chromatography (CH<sub>2</sub>Cl<sub>2</sub>; 90%, 80%, 20% CH<sub>2</sub>Cl<sub>2</sub>/acetone; 90%, 80% CH<sub>2</sub>Cl<sub>2</sub>/MeOH;  
20 MeOH; 95% MeOH/H<sub>2</sub>O, 2.0 L each) to obtain twenty fractions (fractions 1 to 20).  
21 Fraction 8 (11.8 g) was further separated by silica gel flash column chromatography  
22 (CH<sub>2</sub>Cl<sub>2</sub>; 95%, 90%, 80%, 50% CH<sub>2</sub>Cl<sub>2</sub>/acetone; 95% CH<sub>2</sub>Cl<sub>2</sub>/MeOH and MeOH, 2 L  
23 each) to give twenty fractions (fractions 8-1 to 8-20). Fraction 8-6 (2.49 g) was further  
24 separated by ODS flash chromatography (90% MeOH/H<sub>2</sub>O and H<sub>2</sub>O, 2L each) and  
25 eight fractions (fractions 8-6-1 to 8-6-8) were obtained. Fraction 8-6-2 (90.3 mg) was  
26 separated by reverse-phase HPLC (Cosmosil  $\pi$ NAP Waters, 80% MeOH) to afford **1**  
27 (12.6 mg) and **2** (12.3 mg).

28 Compound **1**. Yellow oil; IR (neat) 3400, 2974, 1553, 1400, 1347, 1089 cm<sup>-1</sup>; <sup>1</sup>H and

1 <sup>13</sup>C NMR, see Table 1; ESIMS *m/z* 188 [M-H]<sup>+</sup>; HRESIMS *m/z* 188.1053 [M-H]<sup>+</sup> (calcd.  
2 for C<sub>12</sub>H<sub>14</sub>NO 188.1075).

3  
4 *Bioassay: growth regulating activity against lettuce.*<sup>1-3)</sup> Lettuce seeds were put  
5 on filter paper (Advantec No. 2, φ 55 mm; Toyo Roshi Kaisha, Ltd., Japan), soaked in  
6 distilled water in a Petri dish (φ 60 × 20 mm) and incubated in a growth chamber under  
7 dark at 25 °C for 1 day. Each sample was dissolved in 1 mL of methanol (1, 10<sup>-1</sup>, 10<sup>-2</sup>  
8 and 10<sup>-3</sup> μmol/mL ) and then poured on filter paper (φ 55 mm) in a petri dish (φ 60 ×  
9 20 mm). After the solvent was air-dried, 1mL of distilled water was poured on the  
10 sample-loaded paper or intact filter paper (control). The pre-incubated lettuces (n = 7  
11 in each petri dish) were transferred onto the filter paper and incubated in a growth  
12 chamber under dark at 25 °C for 3 days. The lengths of the hypocotyl and the root were  
13 measured using a ruler.

#### 14 15 **Acknowledgement**

16 This work was partially supported by a Grant-in-Aid for Scientific Research on  
17 Innovative Areas “Chemical Biology of Natural Products” from MEXT (Grant  
18 Number 24102513).

#### 19 20 **References**

- 21 1) Fushimi K, Anzai K, Tokuyama S, Kiriiwa Y, Matsumoto N, Sekiya A, Hashizume  
22 D, Nagasawa K, Hirai H and Kawagishi H, *Tetrahedron*, **68**, 1262-1265 (2012)  
23 2) Wu J, Kobori H, Kawaide M, Suzuki T, Choi J-H, Yasuda N, Noguchi K,  
24 Matsumoto T, Hirai H and Kawagishi H, *Biosci. Biotechnol. Biochem.*, **77**, 1779-1781  
25 (2013)  
26 3) Kobori H, Sekiya A, Yasuda N, Noguchi K, Suzuki T, Choi J-H, Hirai H and  
27 Kawagishi H, *Tetrahedron Lett.*, **54**, 5481-5483 (2013)  
28 4) Garlaschelli L, Pang Z, Sterner O and Vidari G, *Tetrahedron*, **50**, 3571-3574 (1994)

1 Legend to figure

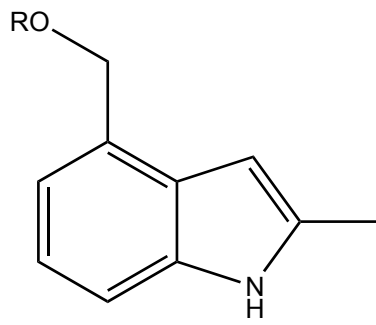
2

3 **Fig. 1.** COSY and HMBC Correlations in **1**.

4

5 **Fig. 2.** Growth Regulating Activity against Lettuce.

6 Black and white columns indicate the elongation of the root and the hypocotyl,  
7 respectively. Each value is presented as the mean  $\pm$  SD of the relative elongation  
8 compared with the control group (n=7). \* $p < 0.01$  (growth inhibition); + $p < 0.01$   
9 (growth promotion).



- 1 R = ethyl
- 2 methyl



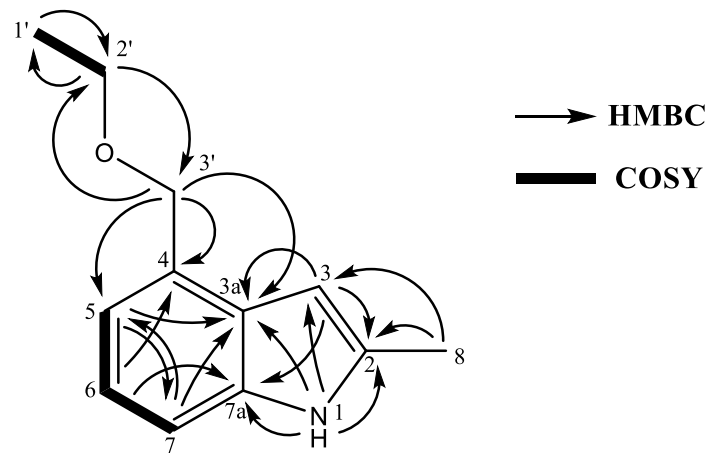


Fig. 1 Qiu et al.

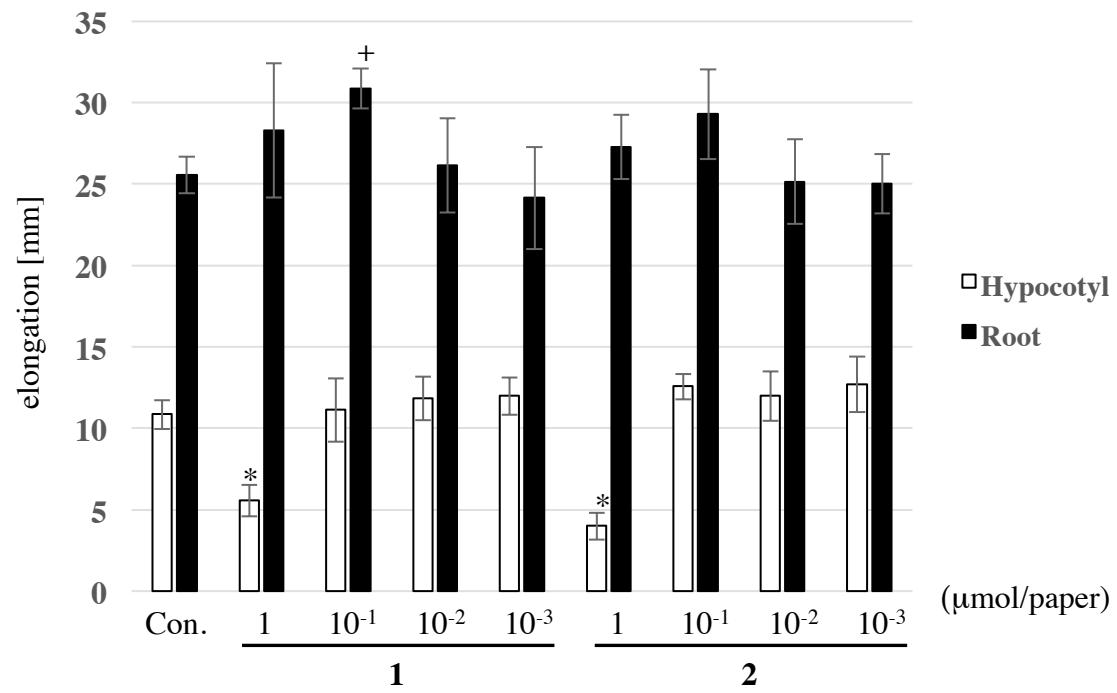


Fig. 2 Qiu et al.

**Table 1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR Data for **1** (in  $\text{CDCl}_3$ )

Position	$^1\text{H}$ ( $\delta$ ; multiplicity; $J$ in Hz)	$^{13}\text{C}$ $\delta$
1	7.88 (br. s)	
2		135.2
3	6.33 (s)	98.0
3a		127.1
4		127.9
5	7.02 (d, 7.3)	118.6
6	7.06 (dd, 7.3, 7.6)	120.1
7	7.21 (d, 7.6)	110.0
7a		135.9
2-Me	2.44 (s)	13.0
4- $\text{CH}_2^-$	4.75 (s)	71.0
$\text{OCH}_2\text{CH}_3$	1.23 (t, 7.0)	14.9
$\text{OCH}_2\text{CH}_3$	3.55 (q, 7.0)	65.2