

A new compound from the mushroom *Tricholoma flavovirens*

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1 Running title: A New Compound from *Tricholoma flavovirens*

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3 **A New Compound from the Mushroom *Tricholoma flavovirens***

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1 A novel compound (**1**) and a known one (**2**) were isolated from the fruiting
2 bodies of *Tricholoma flavovirens*. Their structures were determined by the
3 interpretation of spectroscopic data. Both compounds showed inhibition effects on the
4 growth of hypocotyl of lettuce with significant differences. In addition, compound **1**
5 showed a promotion effect on the growth of root with significant differences and **2** had
6 the similar tendency to promote the growth.

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8 **Key words:** mushroom; *Tricholoma flavovirens*; structural determination; plant growth
9 activity

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11 A well known axiom is that “plants act as producers, animals act as consumers,
12 and fungi act as restorers and decomposers”. Fungi, including mushroom, play an
13 important role in ecological balance as it can restore the nutrients used by plants and
14 animals back to the land. We are interested in biological activity of components from
15 mushroom towards plants and have reported isolation of some compounds that regulate
16 lettuce growth.¹⁻³⁾ Using the assay evaluating growth-regulating activity toward lettuce,
17 we screened extracts of various mushrooms and found relatively strong inhibitory
18 activity in hexane soluble part of the extracts of the mushroom *Tricholoma flavovirens*.

19 Since ancient times *T. flavovirens* (English name, yellow knight; Japanese name,
20 kishimeji) belonging to the family Tricholomataceae is known as an eatable mushroom
21 throughout the world. Till now, few chemical studies were, there, so with the purpose
22 to find novel constituents with activity from *T. flavovirens*, we started this study.

23 Here we describe the isolation, structural determination of a novel compound
24 and a known one from the fruiting bodies of the fungus along with the biological
25 activity of the compounds.

26 Fresh fruiting bodies of *T. flavovirens* were extracted with EtOH and then with
27 acetone. After the solutions were combined and concentrated, they were partitioned
28 between hexane and H₂O, EtOAc and H₂O, and then *n*-BuOH and H₂O. The

1 hexane-soluble part was fractionated by repeated chromatography. As a consequence,
2 two compounds (**1** and **2**) were purified.

3 Compound **1** isolated as yellow oil with a molecular formula determined as
4 $C_{12}H_{15}NO$ by HRESIMS at m/z 188.1053 $[M - H]^-$ (calcd. for $C_{12}H_{14}NO$ 188.1075),
5 indicating presence of six degrees of unsaturation in the molecule. The structure of **1**
6 was elucidated by interpretation of NMR spectra including DEPT, COSY, HMQC, and
7 HMBC (Fig. 1) with the complete assignment of protons and carbons of NMR was
8 accomplished as shown in Table 1. The DEPT experiment indicated the presence of
9 two methyls, two methylenes, four methines and four quaternary carbons. The
10 structure of 2-methylindole was elucidated by the HMBC correlations (H-1/C-2, C-3,
11 C-3a, C-7a; H-3/C-2, C-3a, C-7a; H-5/C-3a, C-7; H-6/C-4, C-7a; H-7/C-3a, C-5;
12 H-8/C-2, C-3) and the COSY correlations (H-5/H-6, H-6/H-7). The HMBC
13 correlations (H-1'/C-2'; H-2'/C-1', C-3'; H-3'/C-2') and the COSY correlations
14 (H-1'/H-2') indicated the presence of the ethoxymethyl group. The connection
15 between the 2-methylindole and ethoxymethyl moiety was confirmed by the HMBC
16 correlations (H-3'/C-3a, C-4, C-5). As a result, the structure **1** was determined as
17 shown.

Fig. 1

Table 1

18 Compound **2** was identified as 4-methoxymethyl-2-methylindole and has been
19 isolated from the fruiting bodies of *Tricholoma sciodes* and *Tricholoma virgatum*.⁴⁾
20 However, no known biological activity of the compound has yet been reported.

21 Both compounds **1** and **2** showed inhibition effects on the growth of hypocotyl
22 of lettuce at 1 μmol /paper with significant differences. In addition, compound **1**
23 showed a promotion effect on the growth of root at 10^{-1} μmol /paper with significant
24 differences and **2** had the similar tendency to promote the growth (Fig. 2). The result
25 indicated that compounds **1** and **2** possessed similar growth regulation activity against
26 lettuce.

Fig. 2

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28

1 *Experimental*

2 *General experiments.* ¹H-NMR spectra (one- and two-dimensional) were recorded
3 on a Jeol lambda-500 spectrometer at 500 MHz, while ¹³C-NMR spectra were recorded
4 by the same instrument at 125 MHz. A JASCO grating infrared spectrophotometer was
5 used to record the IR spectra. The HRESIMS data were measured by a JMS-T100LC
6 mass spectrometer. HPLC separation was performed with a Jasco Gulliver system
7 using a reverse-phase HPLC column (Cosmosil π NAP Waters, Nacalai tesque, Japan).
8 Silica gel plate (Merck F₂₅₄), silica gel 60N (Merck 100-200 mesh), and C₁₈-OPN
9 (Cosmosil 140 μ m) were used for analytical TLC and for flash column
10 chromatography, respectively.

11 *Fungal strain and plant materials.* Mature fruiting bodies of *T. flavovirens*
12 were collected at Narusawa village, Yamanashi Prefecture in Japan. Lettuce seeds
13 (*Lactuca sativa* L. cv. Great Lakes 366; Takii Co., Ltd., Japan) were used in this study.

14 *Extraction and isolation.* The fresh fruiting bodies of *T. flavovirens* (20.6 kg)
15 were extracted with EtOH (42 L, 3 times) and then with acetone (20 L, 3 times). After
16 the solutions were combined and concentrated under reduced pressure, the concentrate
17 was partitioned between hexane and H₂O, EtOAc and H₂O, and then *n*-BuOH and H₂O.
18 The hexane-soluble part (39.8 g) was fractionated by silica gel flash column
19 chromatography (CH₂Cl₂; 90%, 80%, 20% CH₂Cl₂/acetone; 90%, 80% CH₂Cl₂/MeOH;
20 MeOH; 95% MeOH/H₂O, 2.0 L each) to obtain twenty fractions (fractions 1 to 20).
21 Fraction 8 (11.8 g) was further separated by silica gel flash column chromatography
22 (CH₂Cl₂; 95%, 90%, 80%, 50% CH₂Cl₂/acetone; 95% CH₂Cl₂/MeOH and MeOH, 2 L
23 each) to give twenty fractions (fractions 8-1 to 8-20). Fraction 8-6 (2.49 g) was further
24 separated by ODS flash chromatography (90% MeOH/H₂O and H₂O, 2L each) and
25 eight fractions (fractions 8-6-1 to 8-6-8) were obtained. Fraction 8-6-2 (90.3 mg) was
26 separated by reverse-phase HPLC (Cosmosil π NAP Waters, 80% MeOH) to afford **1**
27 (12.6 mg) and **2** (12.3 mg).

28 Compound **1**. Yellow oil; IR (neat) 3400, 2974, 1553, 1400, 1347, 1089 cm⁻¹; ¹H and

1 ¹³C NMR, see Table 1; ESIMS *m/z* 188 [M-H]⁻; HRESIMS *m/z* 188.1053 [M-H]⁻ (calcd.
2 for C₁₂H₁₄NO 188.1075).

3
4 *Bioassay: growth regulating activity against lettuce.*¹⁻³⁾ Lettuce seeds were put
5 on filter paper (Advantec No. 2, φ 55 mm; Toyo Roshi Kaisha, Ltd., Japan), soaked in
6 distilled water in a Petri dish (φ 60 × 20 mm) and incubated in a growth chamber under
7 dark at 25 °C for 1 day. Each sample was dissolved in 1 mL of methanol (1, 10⁻¹, 10⁻²
8 and 10⁻³ μmol/mL) and then poured on filter paper (φ 55 mm) in a petri dish (φ 60 ×
9 20 mm). After the solvent was air-dried, 1mL of distilled water was poured on the
10 sample-loaded paper or intact filter paper (control). The pre-incubated lettuces (n = 7
11 in each petri dish) were transferred onto the filter paper and incubated in a growth
12 chamber under dark at 25 °C for 3 days. The lengths of the hypocotyl and the root were
13 measured using a ruler.

14 15 **Acknowledgement**

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1 Legend to figure

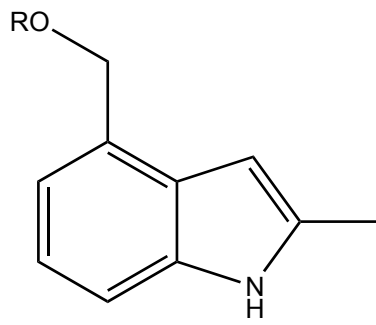
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3 **Fig. 1.** COSY and HMBC Correlations in **1**.

4

5 **Fig. 2.** Growth Regulating Activity against Lettuce.

6 Black and white columns indicate the elongation of the root and the hypocotyl,
7 respectively. Each value is presented as the mean \pm SD of the relative elongation
8 compared with the control group (n=7). * $p < 0.01$ (growth inhibition); + $p < 0.01$
9 (growth promotion).



- 1 R = ethyl
- 2 methyl

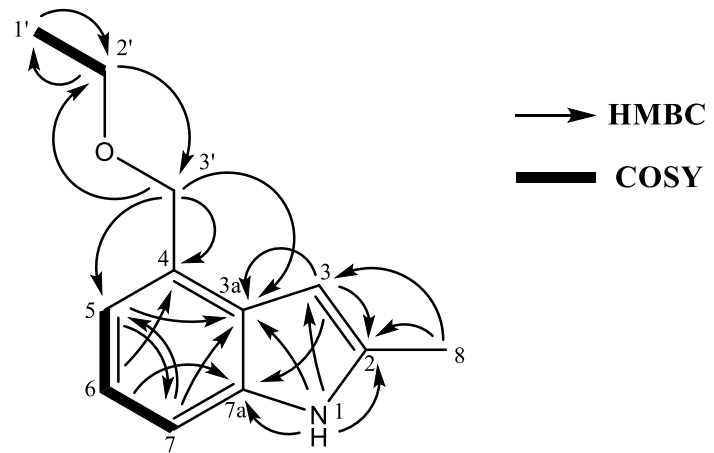


Fig. 1 Qiu et al.

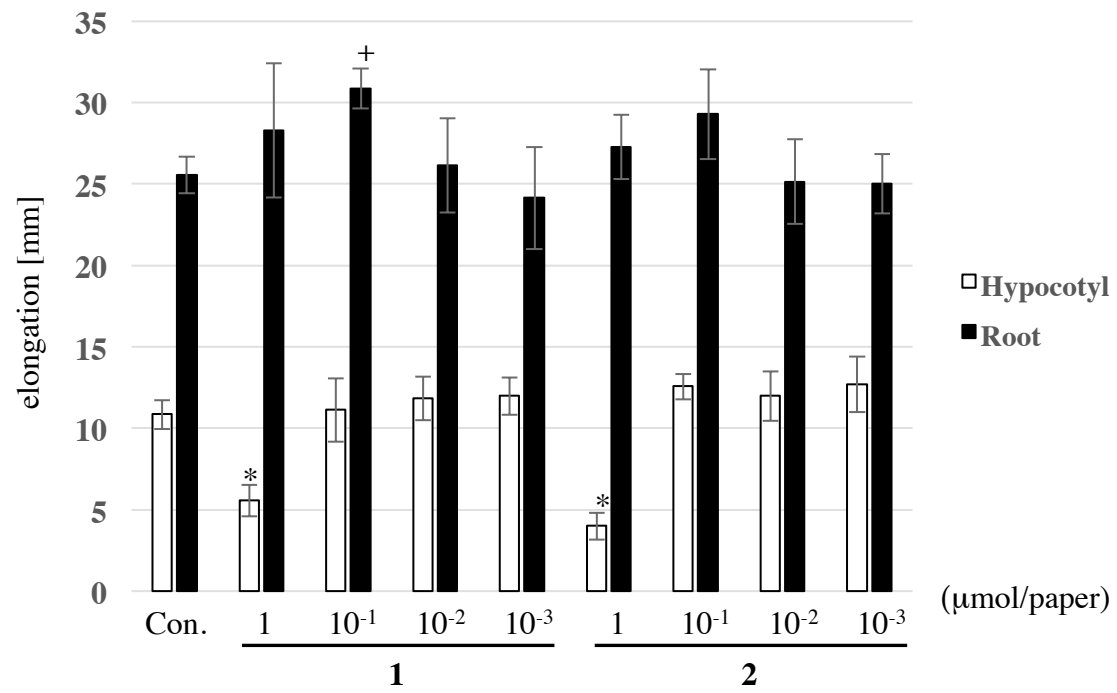


Fig. 2 Qiu et al.

Table 1. ^1H and ^{13}C NMR Data for **1** (in CDCl_3)

Position	^1H (δ ; multiplicity; J in Hz)	^{13}C δ
1	7.88 (br. s)	
2		135.2
3	6.33 (s)	98.0
3a		127.1
4		127.9
5	7.02 (d, 7.3)	118.6
6	7.06 (dd, 7.3, 7.6)	120.1
7	7.21 (d, 7.6)	110.0
7a		135.9
2-Me	2.44 (s)	13.0
4- CH_2^-	4.75 (s)	71.0
OCH_2CH_3	1.23 (t, 7.0)	14.9
OCH_2CH_3	3.55 (q, 7.0)	65.2