

Draft Genome Sequence of *Comamonas testosteronei* R2, Consisting of Aromatic Compound Degradation Genes for Phenol Hydroxylase

メタデータ	言語: eng 出版者: 公開日: 2018-01-11 キーワード (Ja): キーワード (En): 作成者: Azwani, Fatma , Suzuki, Kenshi, Honjyo, Masahiro, Tashiro, Yosuke, Futamata, Hiroyuki メールアドレス: 所属:
URL	http://hdl.handle.net/10297/00024380



Draft Genome Sequence of *Comamonas testosteroni* R2, Consisting of Aromatic Compound Degradation Genes for Phenol Hydroxylase

Fatma Azwani,^{a,c} Kenshi Suzuki,^c Masahiro Honjyo,^b Yosuke Tashiro,^b Hiroyuki Futamata^{b,c,d}

Institute of Tropical Agriculture and Food Security, Universiti Putra Malaysia, Serdang, Selangor, Malaysia^a; Department of Applied Chemistry and Biochemical Engineering, Graduate School of Engineering, Shizuoka University, Hamamatsu, Shizuoka, Japan^b; Graduate School of Science and Technology, Shizuoka University, Hamamatsu, Shizuoka, Japan^c; Research Institute of Green Science and Technology, Shizuoka University, Suruga-ku, Shizuoka, Japan^d

ABSTRACT *Comamonas testosteroni* strain R2 was isolated from a continuous culture enriched by a low concentration of phenol-oxygenating activities with low K_s values (below 1 μM). The draft genome sequence of *C. testosteroni* strain R2 reported here may contribute to determining the phenol degradation gene cluster.

The genome sequencing of many environmental microbes, such as *Comamonas* spp. (1) and *Pseudomonas* spp. (2), has been carried out to better understand the ability of these organisms to use aromatic compounds as sources of carbon. The genome data for these organisms will contribute to our understanding of interspecies interactions and microbial community dynamics (3) and also provide significant insight to the development of bioremediation technologies. Our previous study showed that *C. testosteroni* R2, which was isolated from a chemostat, expressed phenol-oxygenating activities with low apparent K_s (below 1 μM) and had the ability to utilize phenol as a carbon source (4–6).

In order to predict the cluster involved in the expression of the phenol hydroxylase gene of *C. testosteroni* R2, a draft genome sequence was run. The genomic DNA of strain R2 was extracted using a commercial DNA isolation kit, and genome sequencing was performed using a combined method of whole-genome shotgun and paired-end sequencing (7, 8). Draft genome sequence data for strain R2 were generated using 454 GS-FLX Titanium paired-end data (Roche, Basel, Switzerland) (8), which consisted of 631,574 chemistry reads and a total of 189,459,207 bp of sequencing data. The removal of adapter sequences and quality trimming were performed in all data sets prior to *de novo* assembly to correct potential base errors and increase consensus quality. Newbler GS *de novo* assembler version 2.5 software (8) was used to assemble the reads into five scaffolds, with an N_{50} length of 34,745 bp. The draft genome sequence of R2 was estimated to comprise 5,871,018 bp.

The resulting DNA scaffolds, as translational products of coding sequences, were further analyzed by searching the GenBank database to find predicted protein-coding genes, tRNAs, and rRNAs. By using a combination of the Rapid Annotations using Subsystems Technology (RAST) server (9) and the Microbial Genome Annotation Pipeline (MiGAP) (<http://www.migap.org>), the genome was estimated to have an overall G+C content of 60.9%. In total, 5,512 coding regions and 61 tRNAs were predicted and annotated. The rRNAs were further identified using the Southern blot hybridization protocol, which contains five 5S-16S-23S clusters.

Received 14 July 2017 Accepted 19 July 2017 Published 7 September 2017

Citation Azwani F, Suzuki K, Honjyo M, Tashiro Y, Futamata H. 2017. Draft genome sequence of *Comamonas testosteroni* R2, consisting of aromatic compound degradation genes for phenol hydroxylase. *Genome Announc* 5: e00875-17. <https://doi.org/10.1128/genomeA.00875-17>.

Copyright © 2017 Azwani et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Fatma Azwani, fatma@upm.edu.my, or Hiroyuki Futamata, thfutam@ipc.shizuoka.ac.jp.

F.A. and H.F. contributed equally to this article.

The genes encoding phenol hydroxylase of strain R2 were identified. This strain contained one multicomponent phenol hydroxylase and one phenol-metabolic pathway. Under aerobic conditions, phenol hydroxylase is responsible for converting phenol to catechol (10) by incorporating a single hydroxyl group into the substrate—the initial and rate-limiting step in phenol degradation pathways (11). This hydroxylation is followed by ring cleavage that converts catechol by catechol 2,3-dioxygenase C23O (12) to the other metapathway enzymes, such as pyruvate, succinate, and acetyl coenzyme A. Information about the genome sequence of *C. testosteroni* R2 will be helpful for understanding the diversity and mechanisms of phenol degradation in the environment and for furthering bioremediation research.

Accession number(s). The draft genome sequence of strain R2 has been deposited in the DDBJ/EMBL/GenBank database under the accession no. [BDQJ0100001](https://doi.org/10.3389/fmicb.2015.01148) to [BDQJ0100005](https://doi.org/10.3389/fmicb.2015.01148).

ACKNOWLEDGMENT

This work, including the efforts of Hiroyuki Futamata, was supported by grants KAKENHI 15K12228 and KAKENHI (B) 26281038 from the Ministry of Education, Culture, Sports, Science and Technology of Japan.

REFERENCES

1. Aziz FA, Suzuki K, Ohtaki A, Sagegami K, Hirai H, Seno J, Mizuno N, Inuzuka Y, Saito Y, Tashiro Y, Hiraishi A, Futamata H. 2015. Interspecies interactions are an integral determinant of microbial community dynamics. *Front Microbiol* 6:1148. <https://doi.org/10.3389/fmicb.2015.01148>.
2. Suzuki K, Aziz FA, Inuzuka Y, Tashiro Y, Futamata H. 2016. Draft genome sequence of *Pseudomonas* sp. LAB-08 isolated from trichloroethene-contaminated aquifer soil. *Genome Announc* 4(5):e00948-16. <https://doi.org/10.1128/genomeA.00948-16>.
3. Futamata H, Harayama S, Watanabe K. 2001. Diversity in kinetics of trichloroethylene-degrading activities exhibited by phenol-degrading bacteria. *Appl Microbiol Biotechnol* 55:248–253.
4. Futamata H, Harayama S, Watanabe K. 2001. Group-specific monitoring of phenol hydroxylase genes for a functional assessment of phenol-stimulated trichloroethylene bioremediation. *Appl Environ Microbiol* 67:4671–4677. <https://doi.org/10.1128/AEM.67.10.4671-4677.2001>.
5. Watanabe K, Teramoto M, Futamata H, Harayama S. 1998. Molecular detection, isolation, and physiological characterization of functionally dominant phenol-degrading bacteria in activated sludge. *Appl Environ Microbiol* 64:4396–4402.
6. Watanabe K, Hino S, Onodera K, Kajie S, Takahashi N. 1996. Diversity in kinetics of bacterial phenol-oxygenating activity. *J Ferment Bioeng* 81: 560–563. [https://doi.org/10.1016/0922-338X\(96\)81481-4](https://doi.org/10.1016/0922-338X(96)81481-4).
7. Rutherford K, Parkhill J, Crook J, Horsnell T, Rice P, Rajandream MA, Barrell B. 2000. Artemis: sequence visualization and annotation. *Bioinformatics* 16:944–945. <https://doi.org/10.1093/bioinformatics/16.10.944>.
8. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen YJ, Chen Z, Dewell SB, Du L, Fierro JM, Gomes XV, Godwin BC, He W, Helgesen S, Ho CH, Ho CH, Irzyk GP, Jando SC, Alenquer ML, Jarvie TP, Jirage KB, Kim JB, Knight JR, Lanza JR, Leamon JH, Lefkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhijani VB, McDade KE, McKenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JF, Simpson JW, Srinivasan M, Tartaro KR, Tomasz A, Vogt KA, Volkmer GA, Wang SH, Wang Y, Weiner MP, Yu P, Begley RF, Rothberg JM. 2005. Genome sequencing in micro-fabricated high-density picolitre reactors. *Nature* 437:376–380. <https://doi.org/10.1038/nature03959>.
9. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
10. Arai H, Akahira S, Ohishi T, Maeda M, Kudo T. 1998. Adaptation of *Comamonas testosteroni* TA441 to utilize phenol: organization and regulation of the genes involved in phenol degradation. *Microbiology* 144:2895–2903. <https://doi.org/10.1099/00221287-144-10-2895>.
11. Hino S, Watanabe K, Takahashi N. 1998. Phenol hydroxylase cloned from *Ralstonia eutropha* strain E2 exhibits novel kinetic properties. *Microbiology* 144:1765–1772. <https://doi.org/10.1099/00221287-144-7-1765>.
12. Arai H, Akahira S, Ohishi T, Kudo T. 1999. Adaptation of *Comamonas testosteroni* TA441 to utilization of phenol by spontaneous mutation of the gene for a trans-acting factor. *Mol Microbiol* 33:1132–1140. <https://doi.org/10.1046/j.1365-2958.1999.01554.x>.